

Bioprocess intensification: Challenges related to transfer limitation

2nd workshop at CRDB/SBFT-HUST (Hanoi, VN)

27-30th June 2016

INVESTIGATION OF PHYSICAL MECHANISMS DURING DECONSTRUCTION OF PRETREATED LIGNOCELLULOSIC MATRIXES: FROM PURE ENZYMATIC ACTIVITY TO COCKTAIL

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LABORATOIRE D'INGÉNIERIE
DES SYSTÈMES BIOLOGIQUES
ET DES PROCÉDÉS



- Enzymatic hydrolysis.
- Physical analysis in-situ/offline (rheo-morphology-granulo, kinetic of decantation,...)

- Chemical analysis (intermediaries of degradation, composition, cristallinity..._



IMFT
(Toulouse)

LISBP
(Toulouse)

LCPO
(Bordeau)



- Physical approach: rheology of suspension

WU
(Canada)

SBFT
(Hanoi)

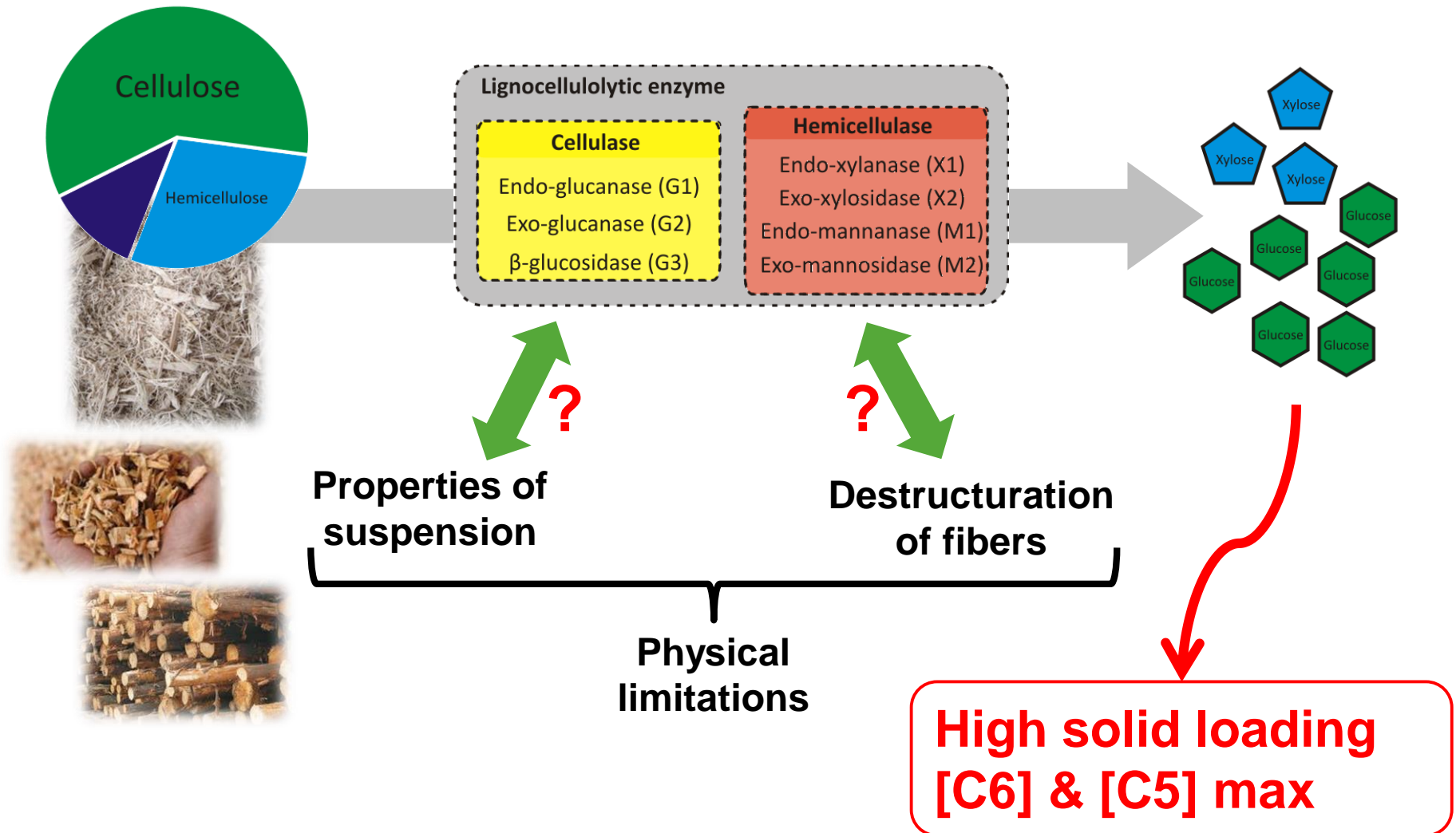
- Analysis of surface's properties.

- Thermo-chemical pretreatment of biomass.
- Biochemical analysis.

Financial Supports:

Erasmus Mundus - Techno 1
BioAsie – HTMS

Enzymatic hydrolysis of lignocellulosic biomass



Physical approach

- Properties of lignocellulosic fiber suspension (yield stress, rheological behavior, particle size and morphology)
- Evolution of fiber properties during enzymatic hydrolysis

Knowledge

-Lignocellulosic suspension possess **shear thinning properties** (non-Newtonian fluids which have decreased viscosity when subjected to shear strain)

μ , P = f (solid loading) : 4% increase in solid content lead to 5 folds rise in power consumption (Fan et al., 2003; Dunaway et al. 2010; Knutsen et al. 2012)

μ = f (particle size) : at 10% w/w, 50 folds rise in viscosity from particle size 33-75 μm up to 150-180 μm (Dasari and Berson 2007)

Role of **single activities** in the liquefaction : endo-glucanase =dominant (Szijártó et al., 2011)

Physical approach on enzymatic hydrolysis

Physical approach

- Properties of lignocellulosic fiber suspension (yield stress, rheological behavior, particle size and morphology)

Knowledge

- Lignocellulosic suspension possess **shear thinning properties** (non-Newtonian fluids which have decreased viscosity when subjected to shear strain)

$\mu, P = f(\text{solid loading})$: 4% increase in solid content lead to 5 folds rise in power consumption (Fan et al., 2003)

Lack in knowledge

Investigation of pure up to cocktail activities by physical approach: in-situ viscosity, mechanisms of degradation.

→ My PhD

Substrate

Sugarcane bagasse: from sugar factory Nong Cong and Lam Son, VietNam. Organosolv pretreated + extruded.

Filter paper : Whatman n°1, milled

Hard wood paper pulp: from French industrial.

Enzyme

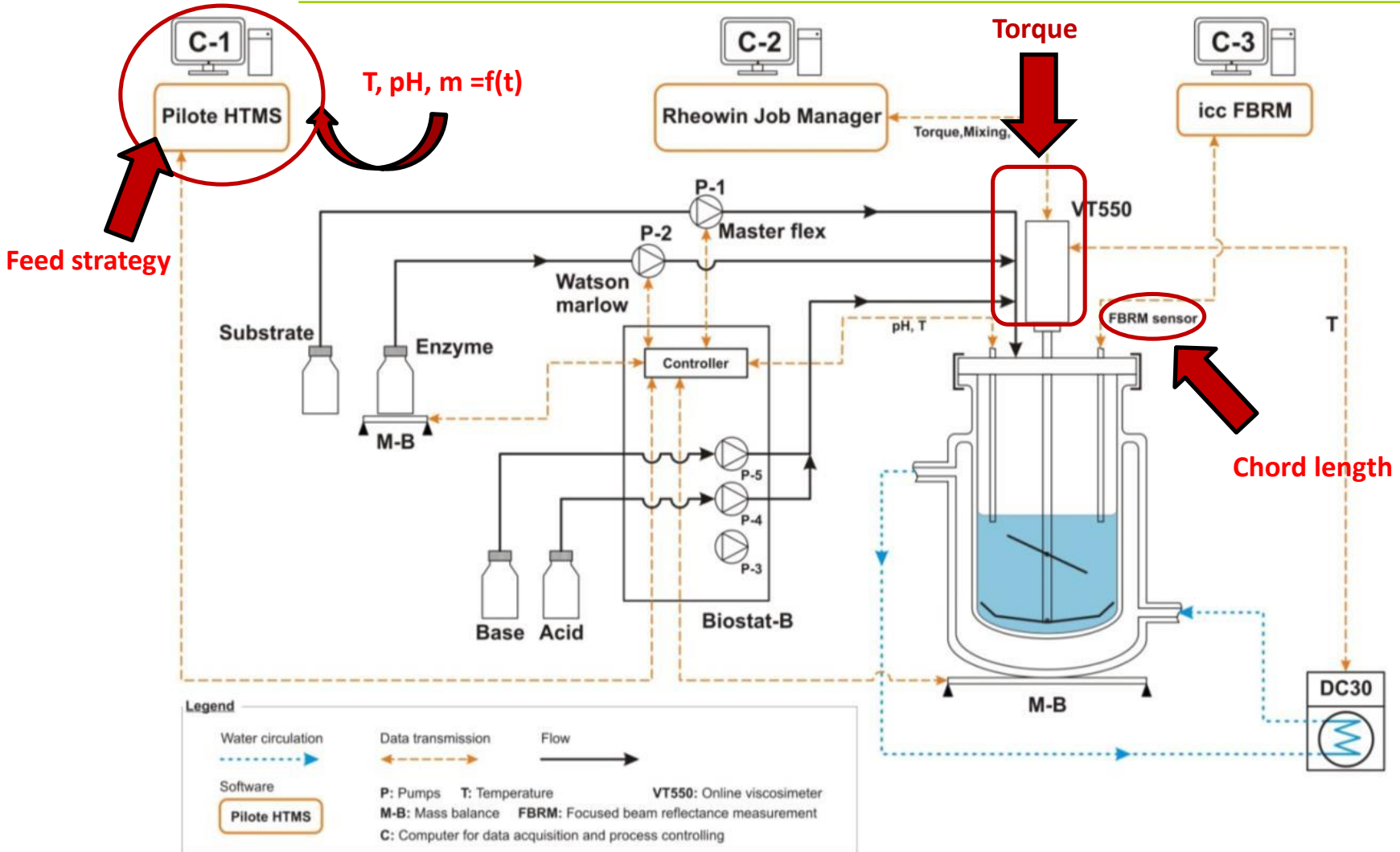
Cellic Ctec2, Novozymes, activity 103FPU/mL at 40°C, pH 4.8

Endo-glucanase: E-CELAN, Megazymes (high purity) – **G1**

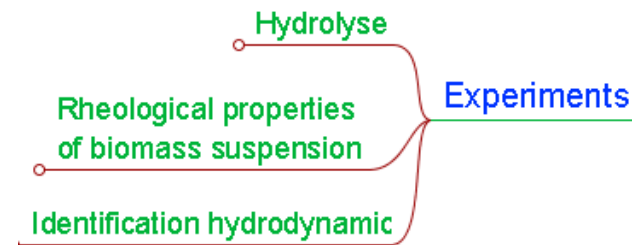
Exo-glucanase: E6412, Sigma – **G2**

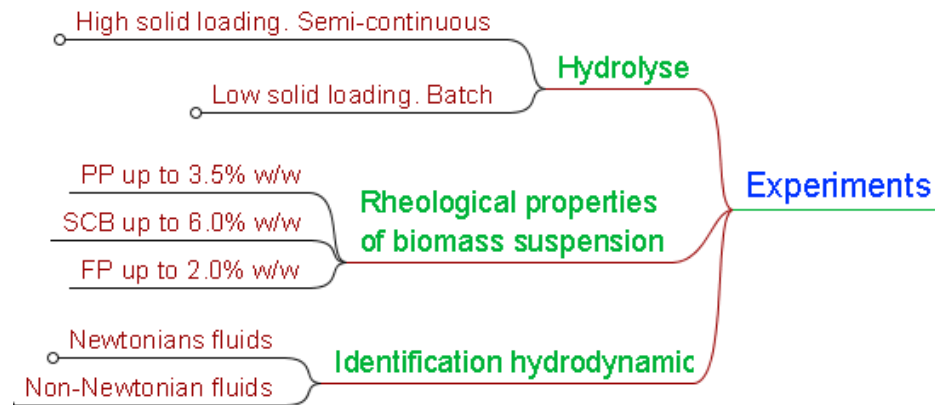
β-glucosidase: 49290, Sigma – **G3**

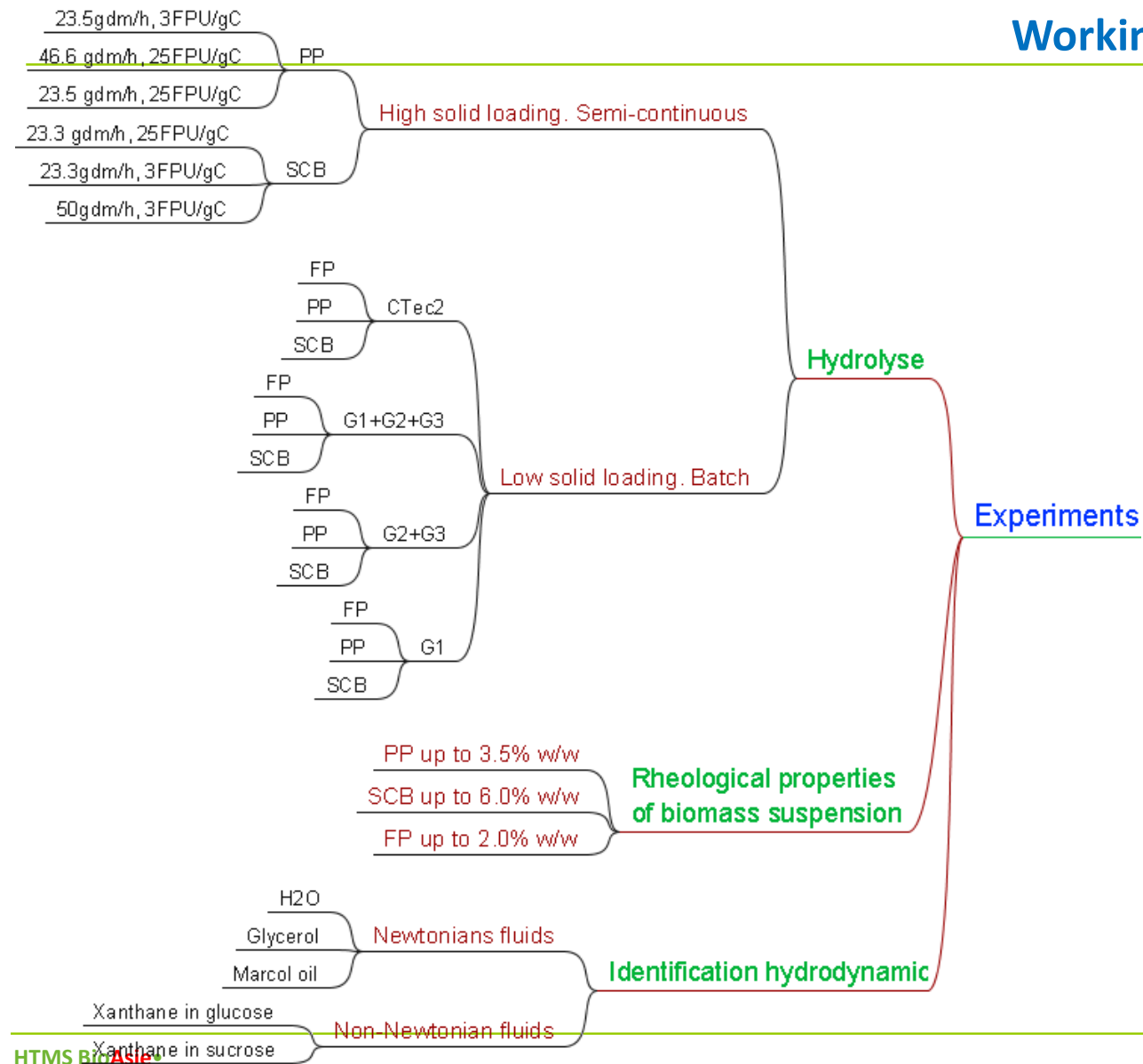




Measurement	Raw data	Interpreted data
In-situ Torque	$M=f(t)$	Viscosity, rheological behavior
In-situ Chord Length	$En(cl), N(cl)=f(t)$	Ev, Fv, En, Fn, dSE
Ex-situ rheology	G', G''	Viscosity, yield stress
DLS	$Ev(dSE), d[4,3]$	$Ev(dSE), Fv(dSE), d[4,3]$
Morphologi	Particle sharpe & size	$Ev(dCE),$ Particle sharpe & size
Decantation kinetic	Settling velocity	
HPLC	Mono to di-saccharides	Hydrolysis yield



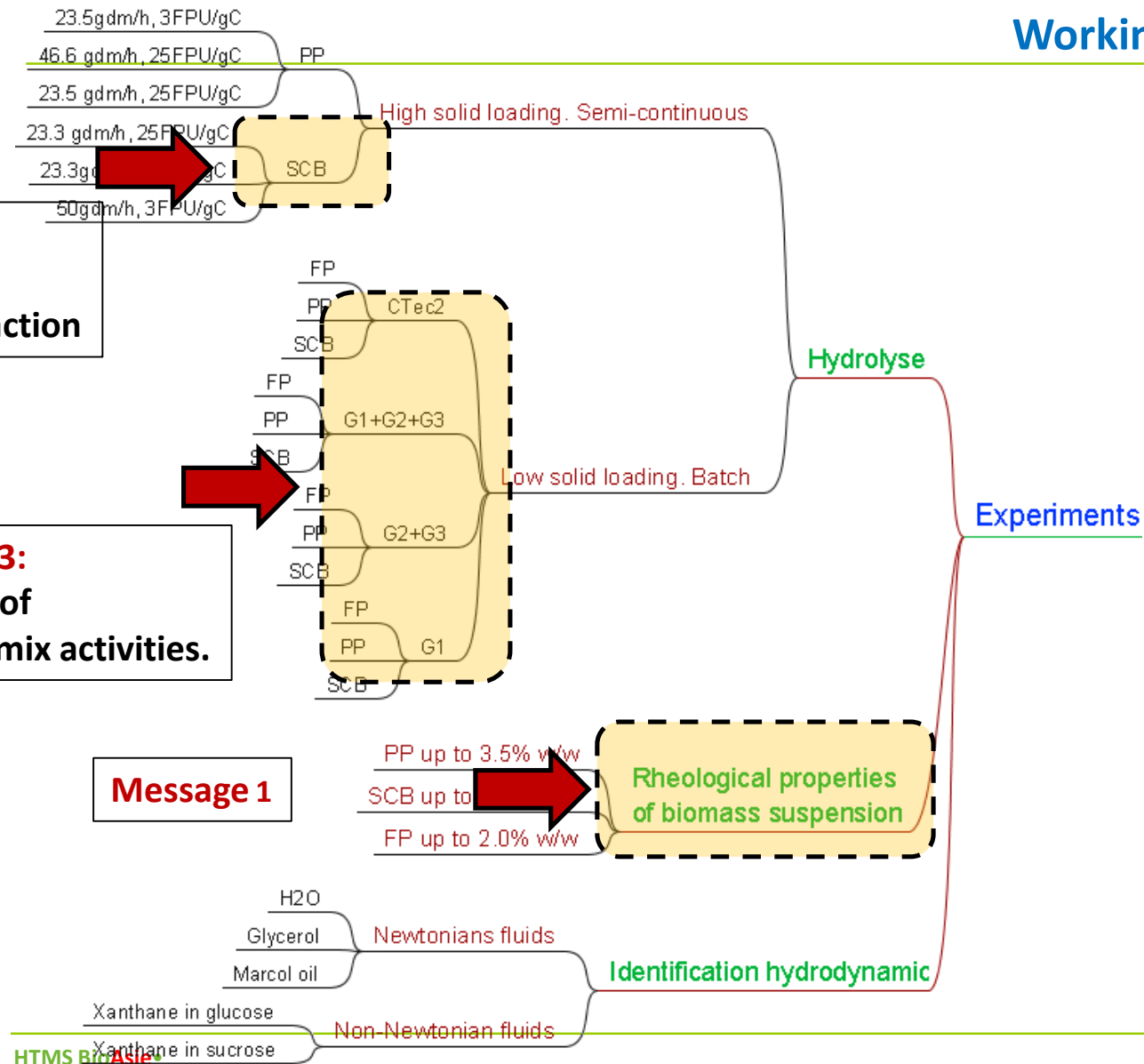


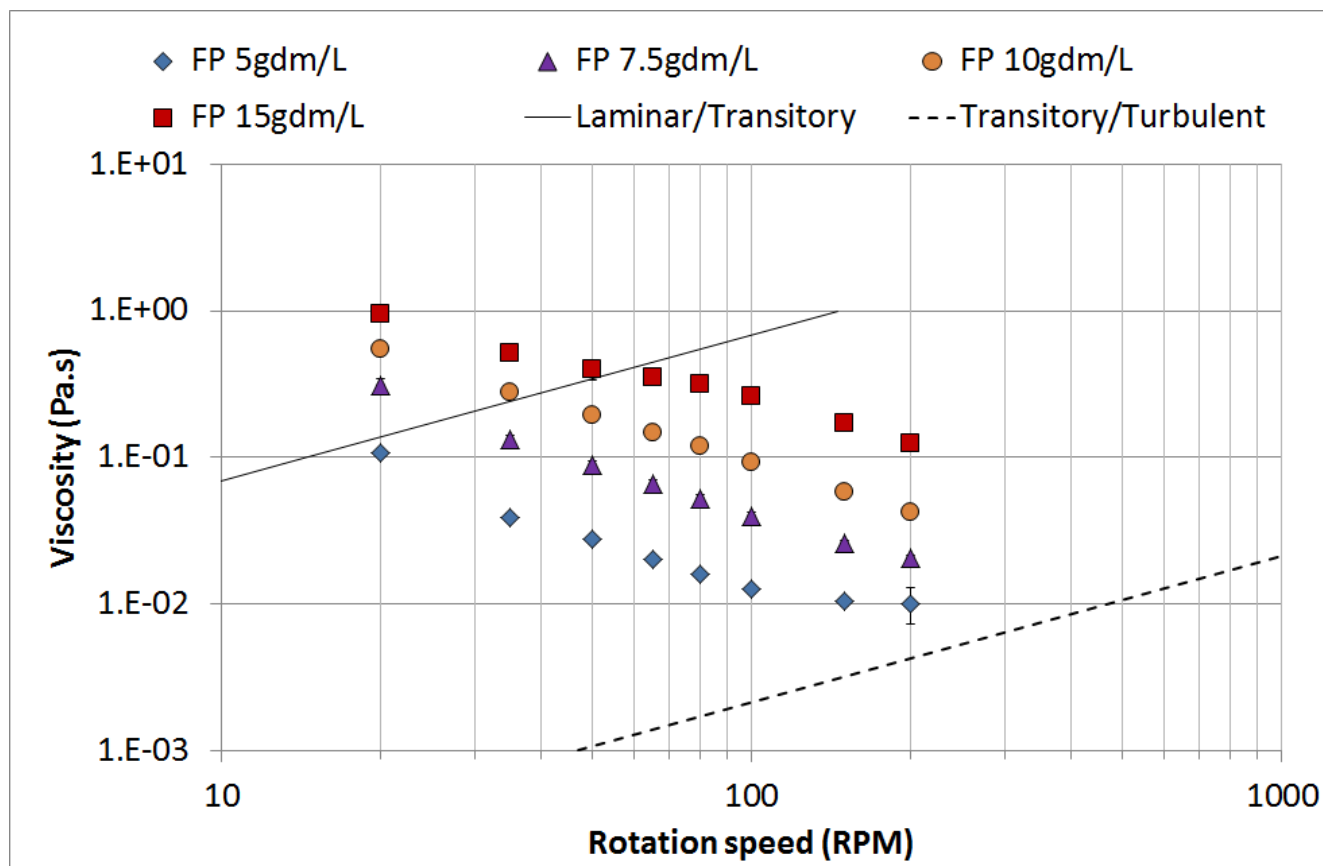


Message 4:
Mechanism of
enzymatic liquefaction

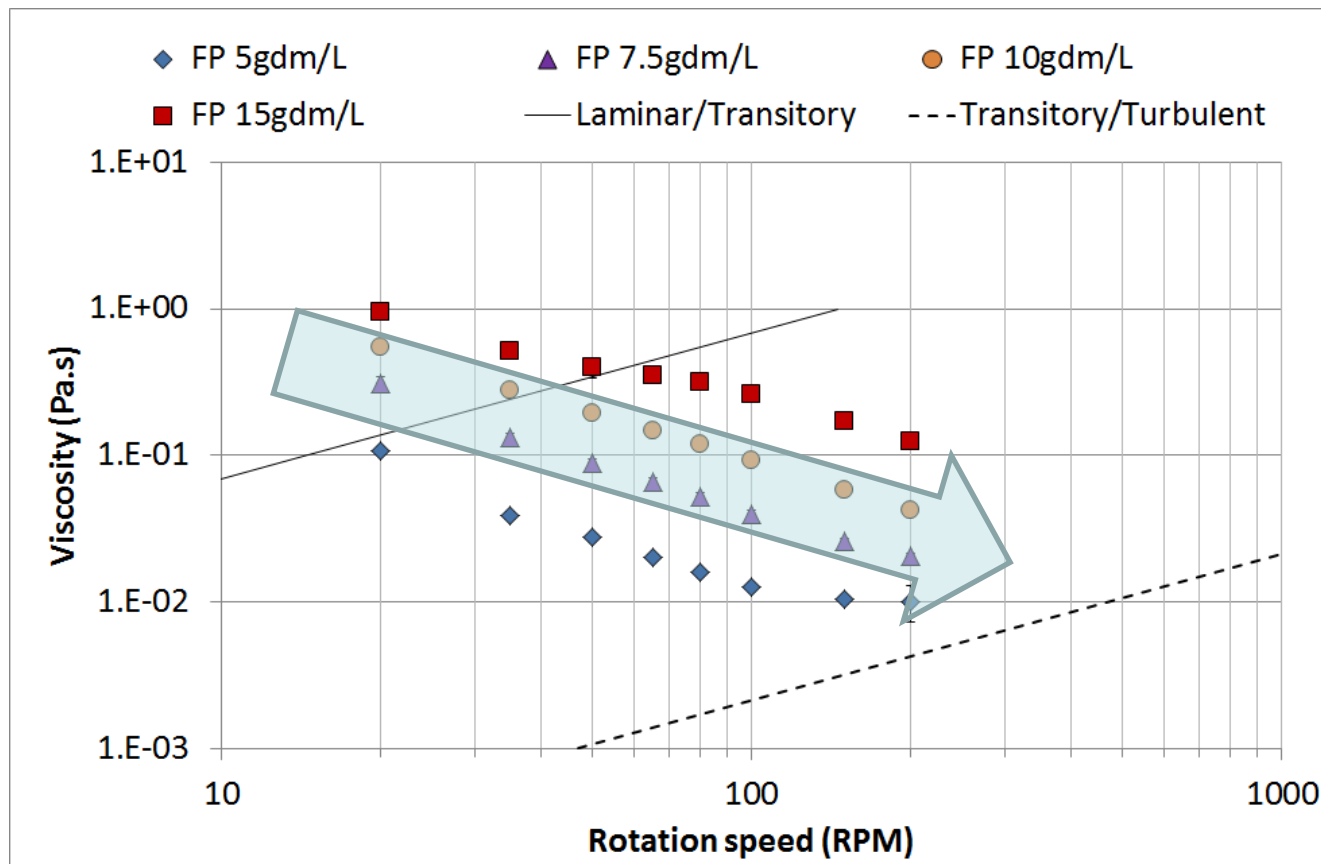
Message 2 & 3:
Investigation of
individual to mix activities.

Message 1





Suspension viscosity at different mixing rate
Substrate: **filter paper**

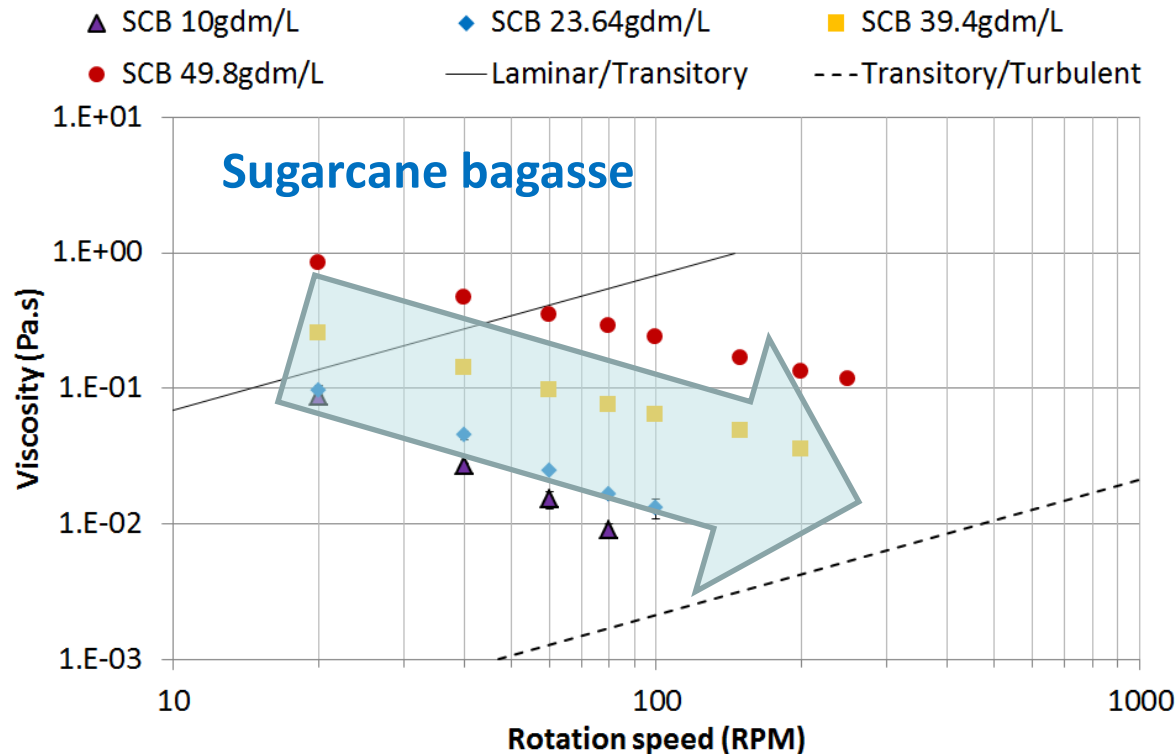


Suspension viscosity at different mixing rate

Substrate: **filter paper**

Suspension viscosity decreased as the mixing rate increased
-> shear-thinning properties

Substrate properties – SCB and PP



Suspension viscosity at different mixing rate
Substrate: **filter paper**

Suspension viscosity decreased as the mixing rate increased
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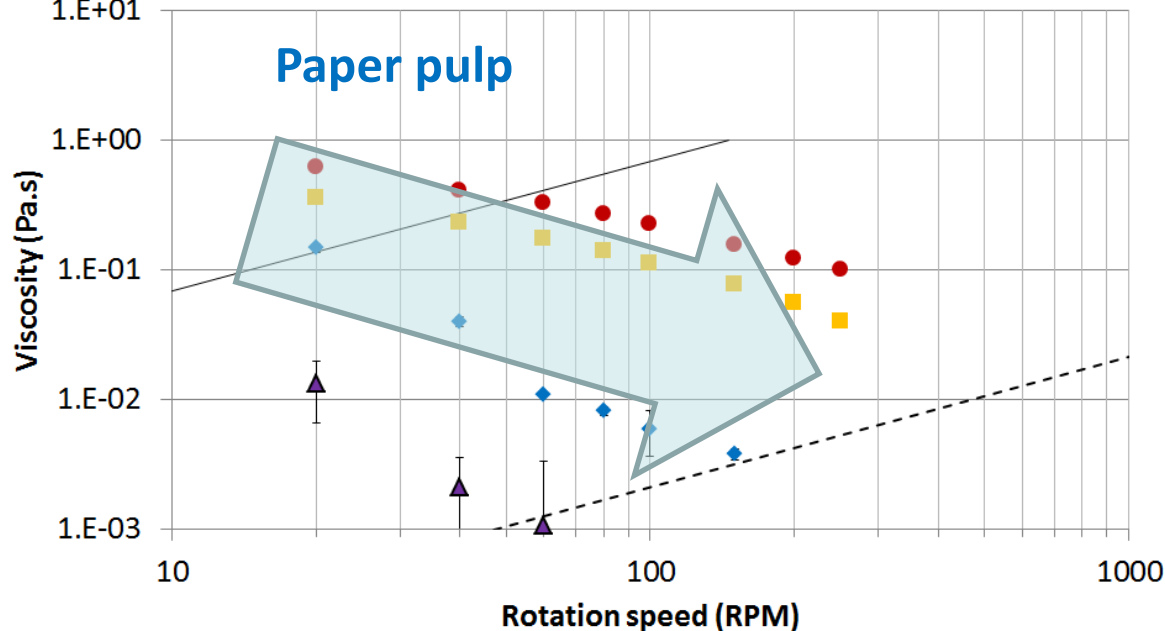
Similar behavior on **sugarcane bagasse** ...

Substrate properties – SCB and PP

▲ SCB 10gdm/L ◆ SCB 23.64gdm/L ■ SCB 39.4gdm/L
 ● SCB 49.8gdm/L — Laminar/Transitory --- Transitory/Turbulent
 1.E+01

Sugarcane bagasse

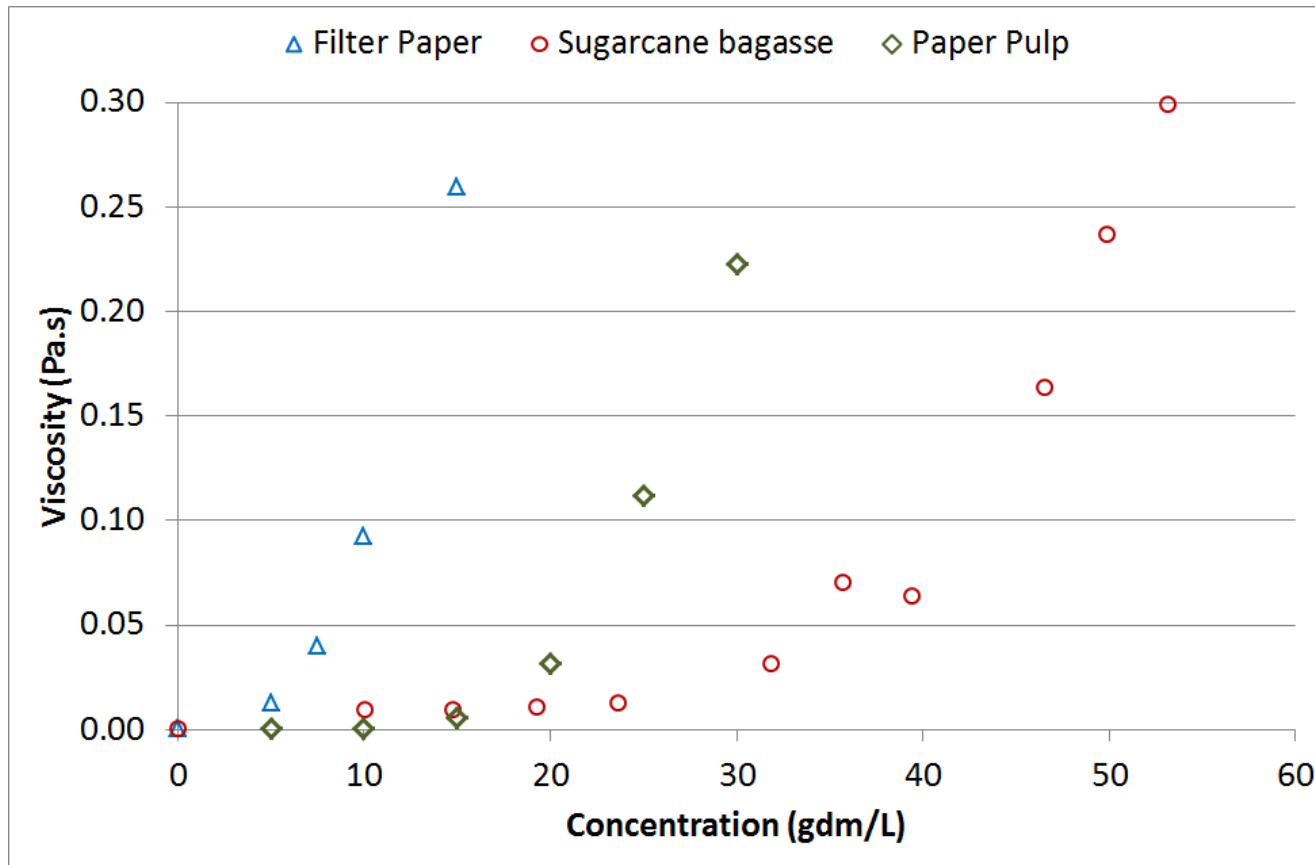
▲ PP 5gdm/L ◆ PP 15gdm/L ■ PP 25gdm/L
 ● PP 30gdm/L — Laminar/Transitory --- Transitory/Turbulent
 1.E+01



Suspension viscosity at different mixing rate
Substrate: **filter paper**

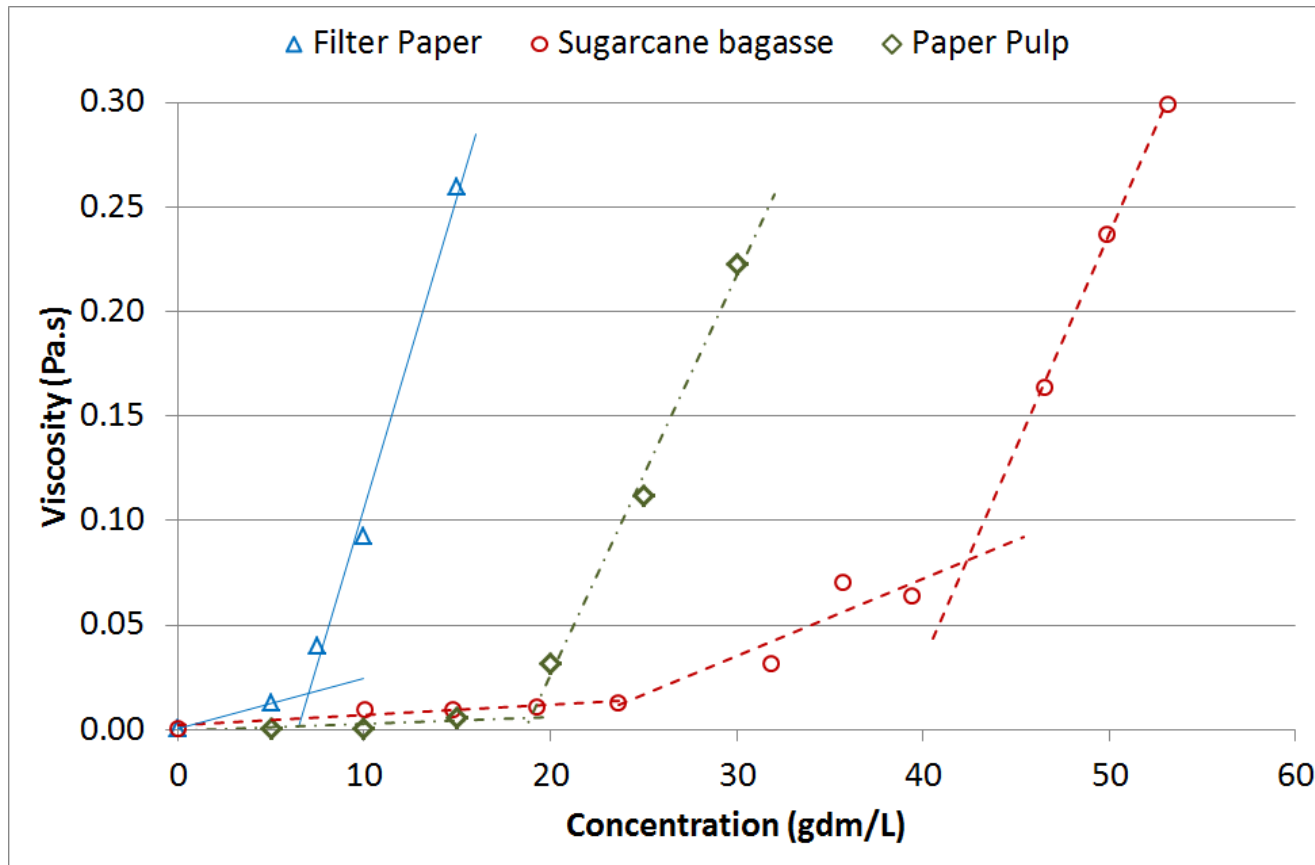
Suspension viscosity decreased as the mixing rate increased
-> shear-thinning properties

Similar behavior on **sugarcane bagasse ...**
and on **paper pulp**



Viscosity at 100 rpm of SCB, FP and PP suspensions in relation with **biomass concentration**

Substrate properties

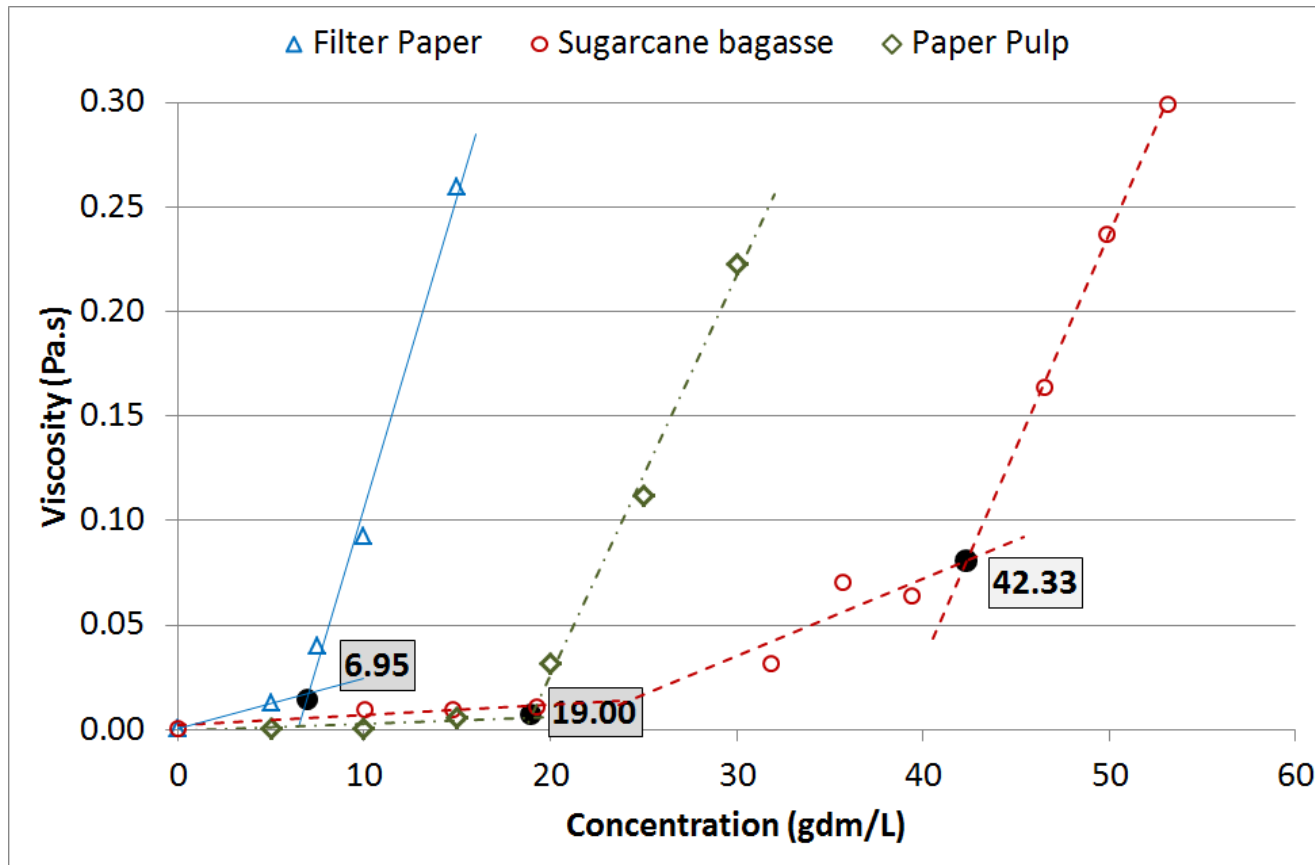


Viscosity at 100 rpm of SCB, FP and PP suspensions in relation with **biomass concentration**



Viscosity rise as **biomass concentration** increased

Substrate properties



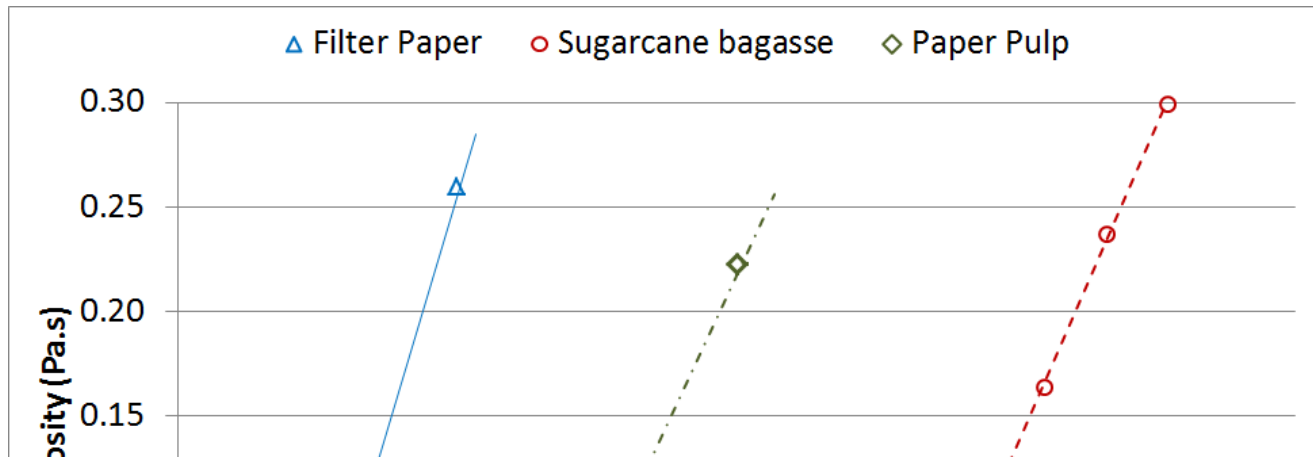
Viscosity at 100 rpm of SCB, FP and PP suspensions in relation with **biomass concentration**



Viscosity rise as **biomass concentration** increased



Identification of substrate's **critical concentration**



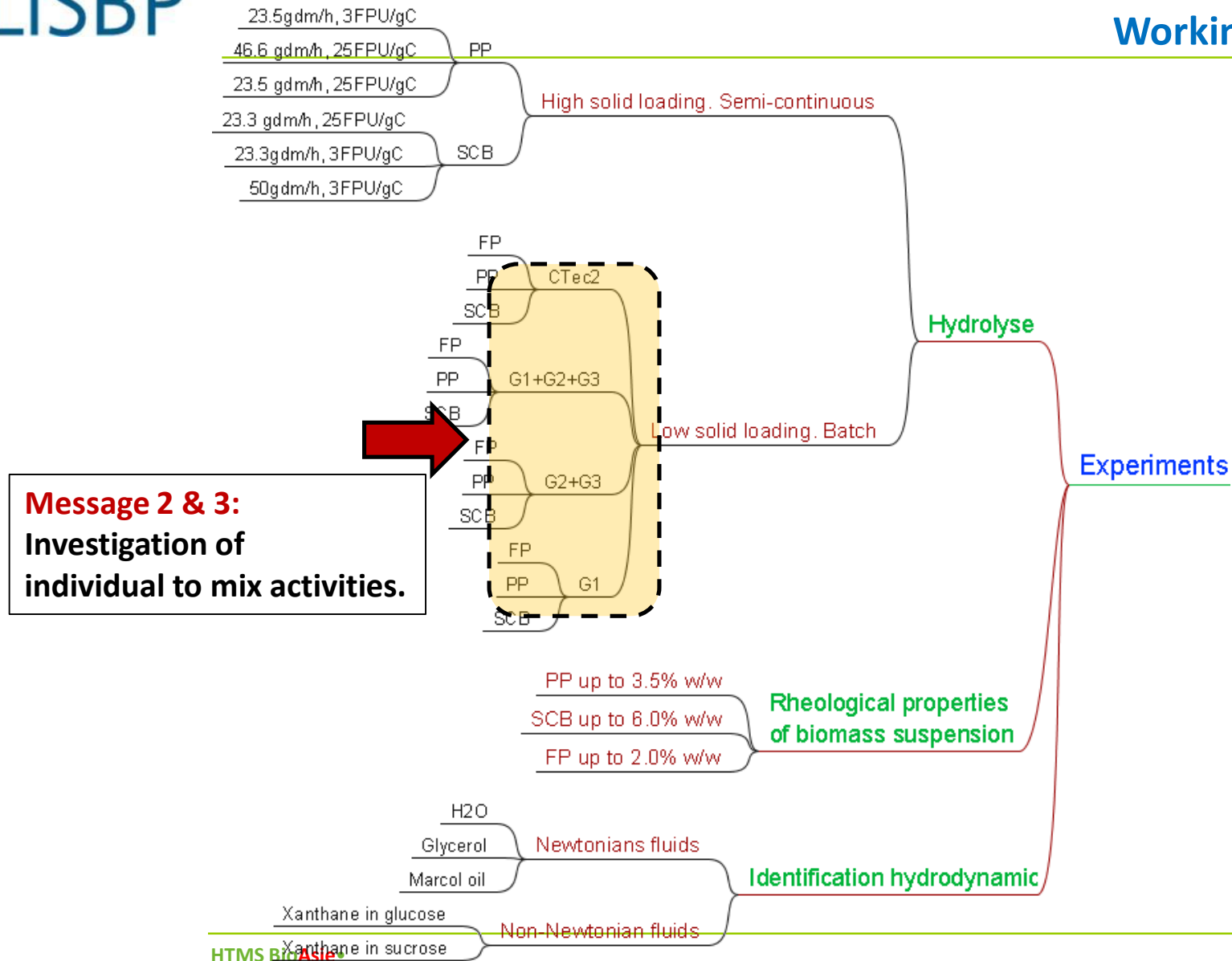
Viscosity at 100 rpm of SCB, FP and PP suspensions in relation with **biomass concentration**



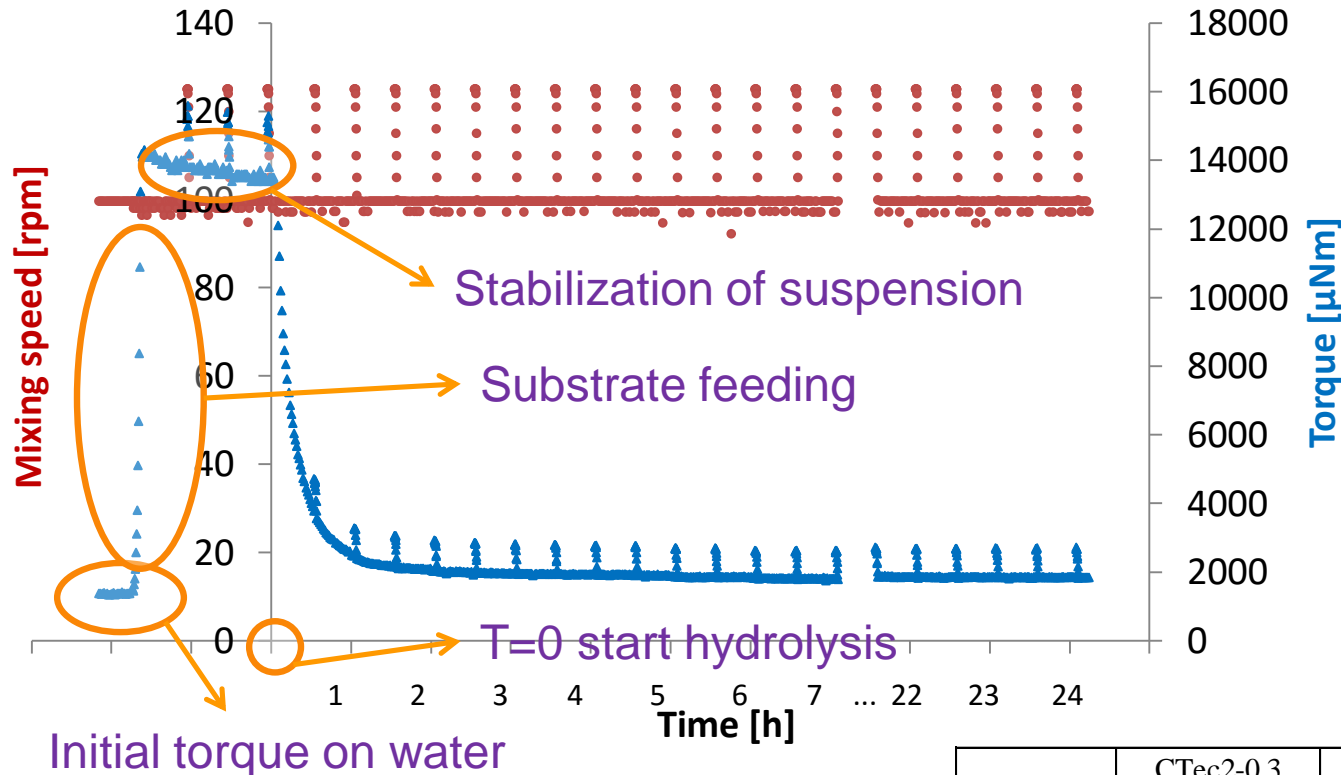
Viscosity rise as **biomass concentration** increased

Message 1

- All studied suspension behaved as shear-thinning fluid.
- Suspension viscosity = $f(\text{solid loading})$.
- Identification of critical concentration points.



• \ddagger in 1/min \blacktriangle M in μNm



Hydrolysis parameters

- Mixing speed:
 - 100 rpm for 28 min
 - 125 rpm for 1 min
 - speed-down to 100 rpm
 - repeat until the end
- pH: 4,8
- Temperature: 40°C
- Substrate: 1,5-3% w/v

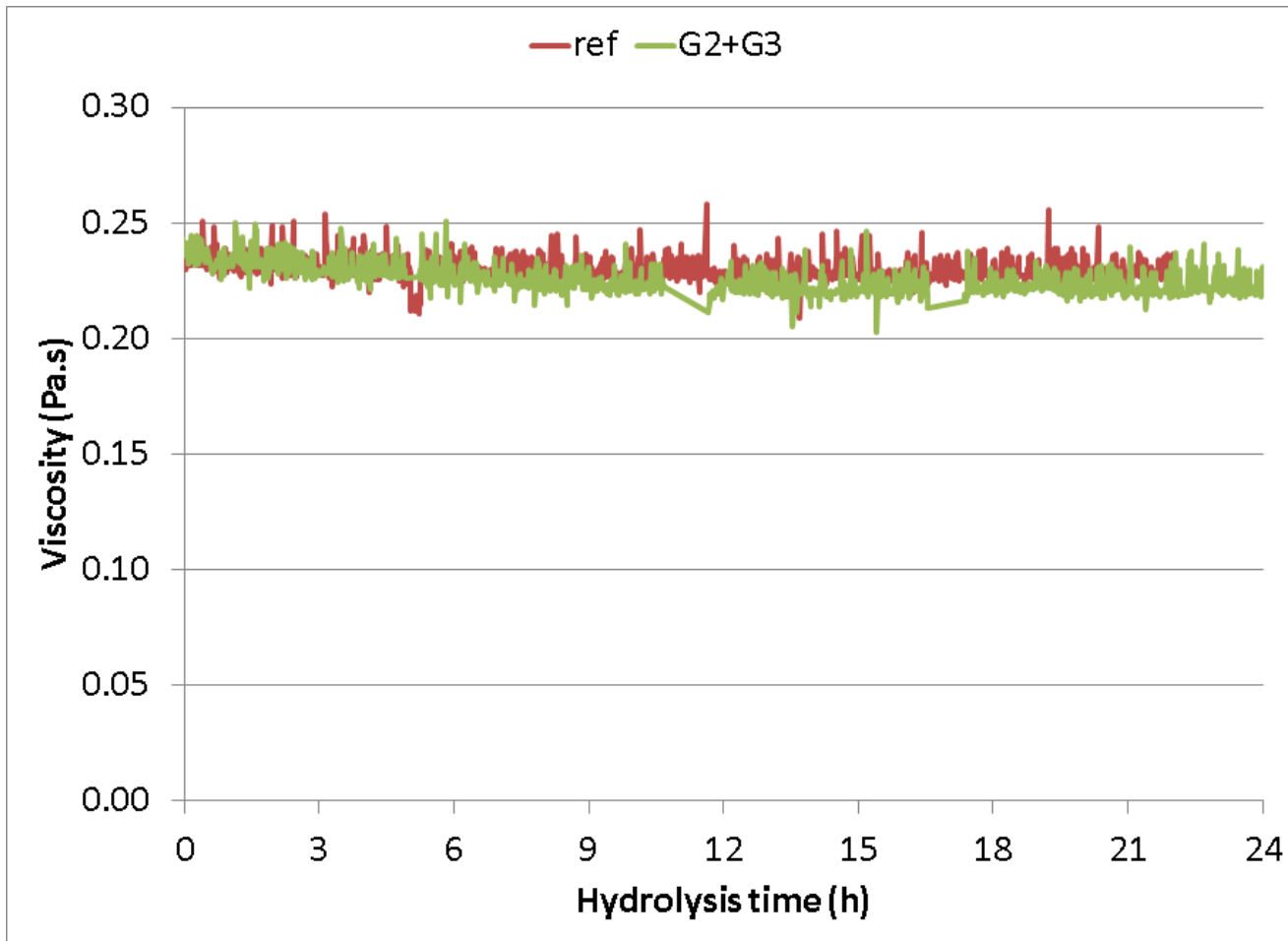
Enzyme dosage

	CTec2-0.3	G1	G2+G3	G1+G2+G3
FPU	0.30	/	/	
CMCU	2.42	2.42	/	2.42
AVCU	0.30	/	3.00	3.00
CBU	10.95	/	0.50	0.50

Sampling at 0-1-2-3-6-12-18-24h

Pure activities contribution for liquefaction

Filter paper 15gdm/L



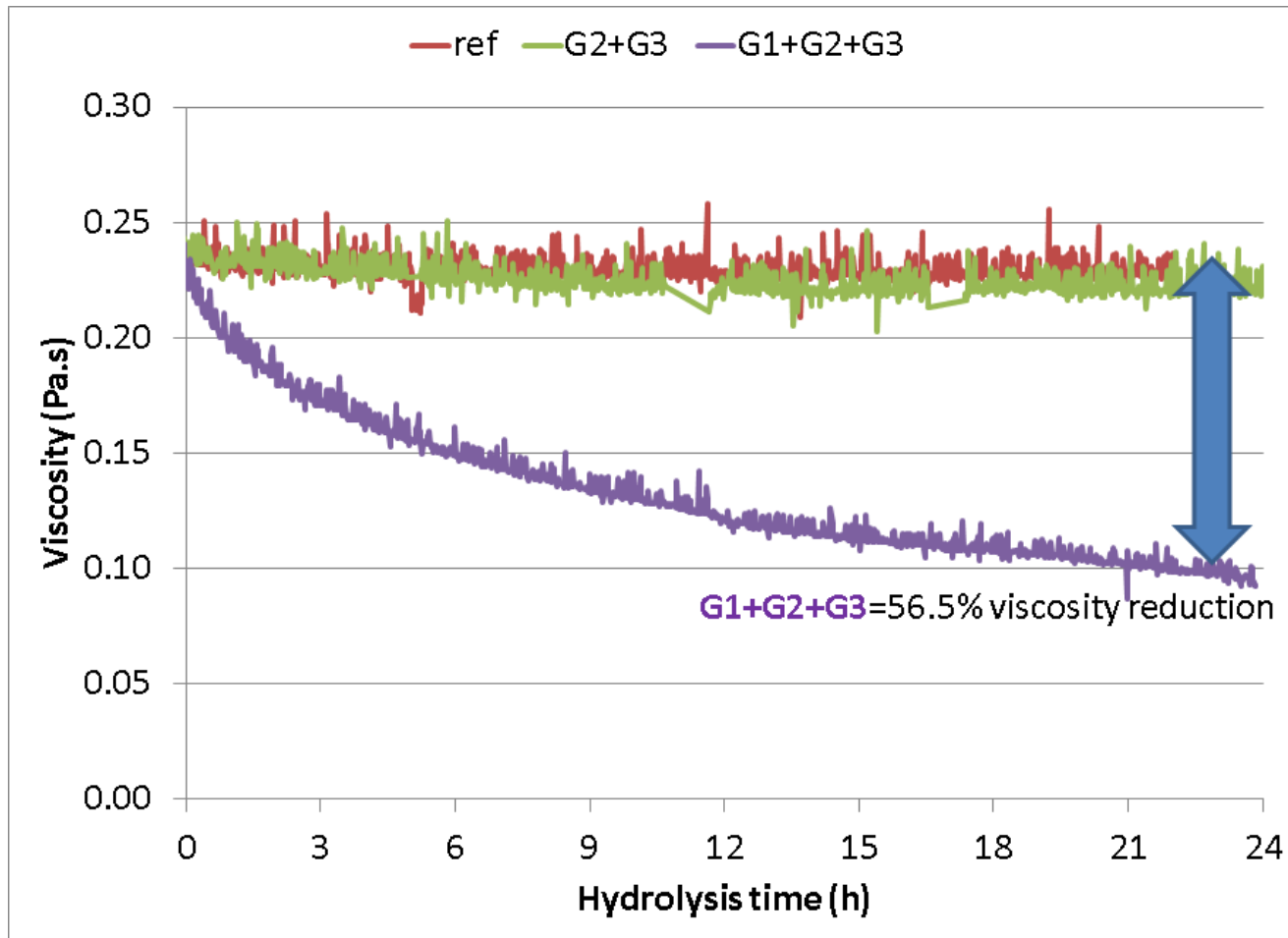
Mixing rate 100rpm

Ref: no enzyme

G2+G3 (exo-glucanase & β -glucosidase) = **no contribution**

Pure activities contribution for liquefaction

Filter paper 15gdm/L



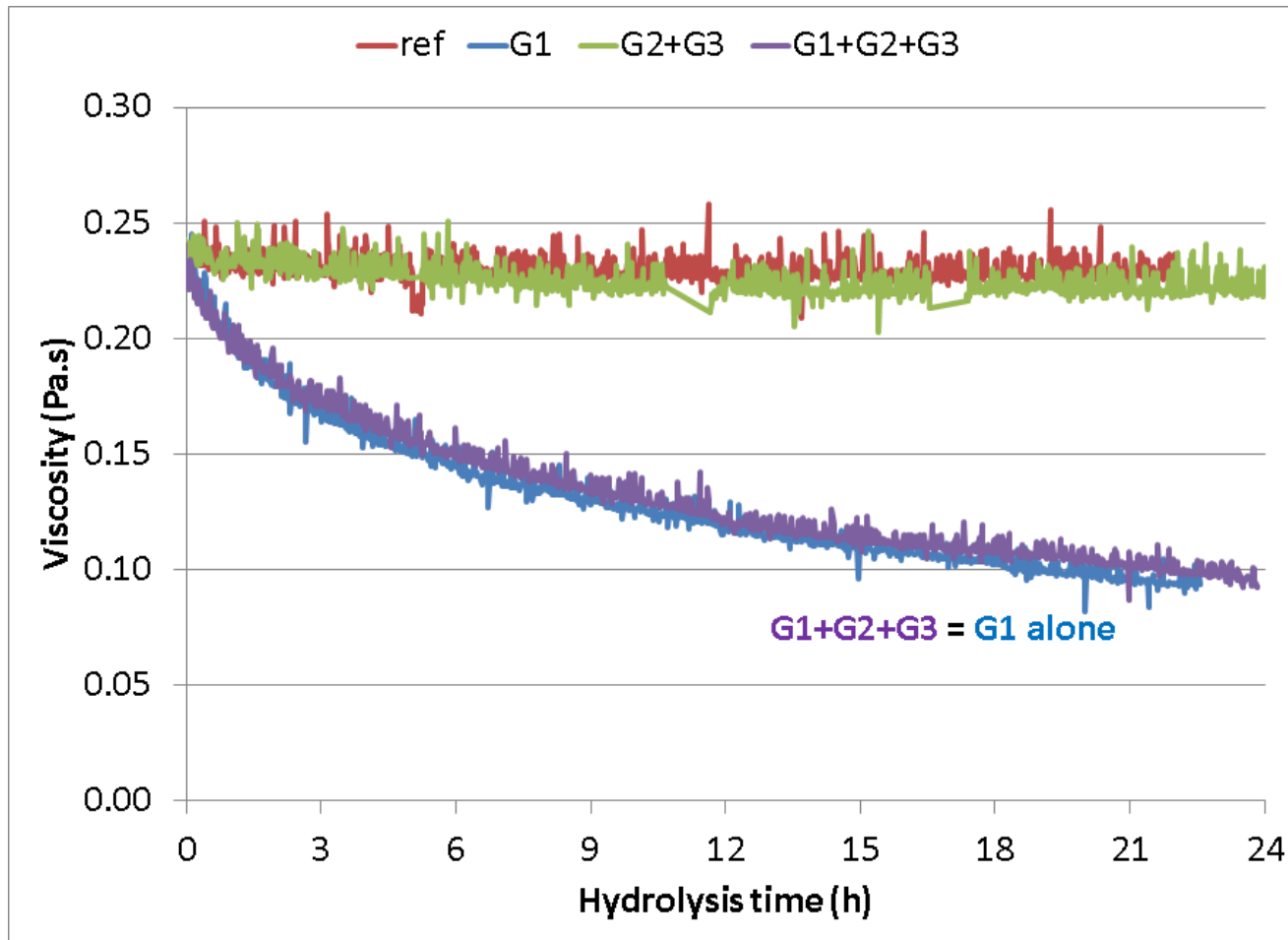
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Pure activities contribution for liquefaction

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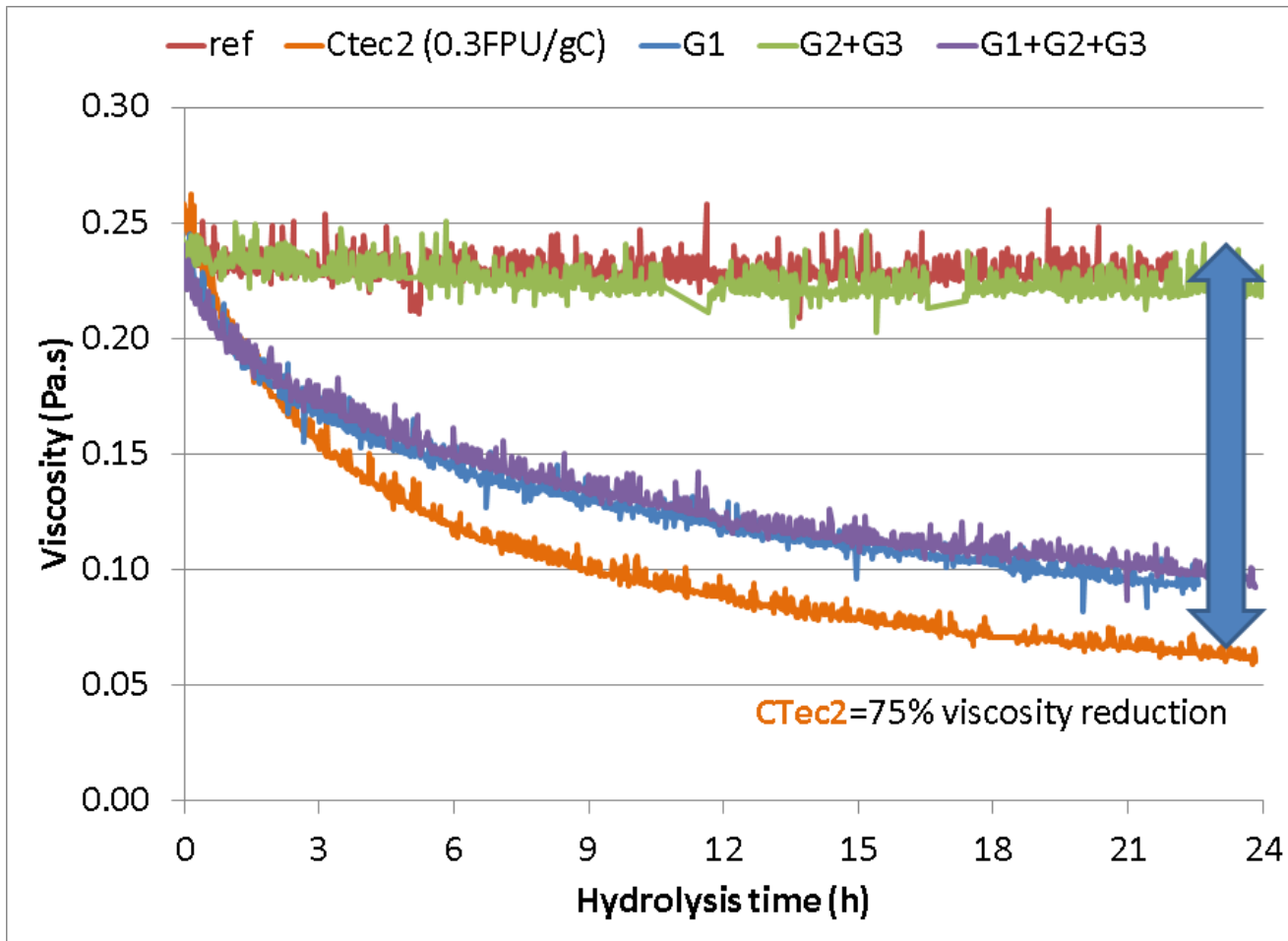
G2+G3 (exo-glucanase & β -glucosidase) = **no contribution.**



G1 (endo-glucanase) = predominant role.

Pure activities contribution for liquefaction

Filter paper 15gdm/L



Mixing rate 100rpm

Ref: no enzyme

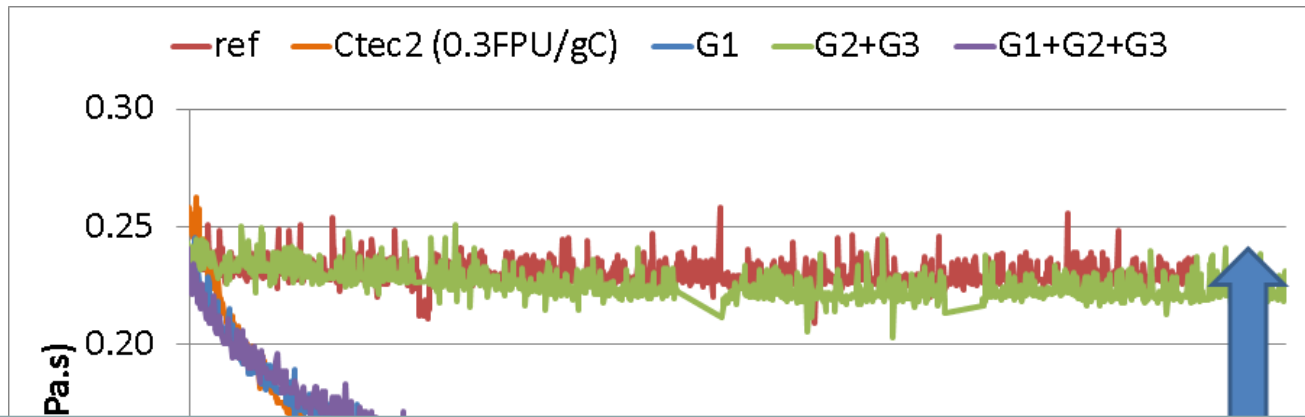
G2+G3 (exo-glucanase & β -glucosidase) = **no contribution**.

G1 (endo-glucanase) = predominant role.

CTec2 = better performance !
Enzyme synergist ?.

Pure activities contribution for liquefaction

Filter paper 15gdm/L



Mixing rate 100rpm

Ref: no enzyme

G2+G3 (exo-glucanase & β -glucosidase) = **no contribution.**

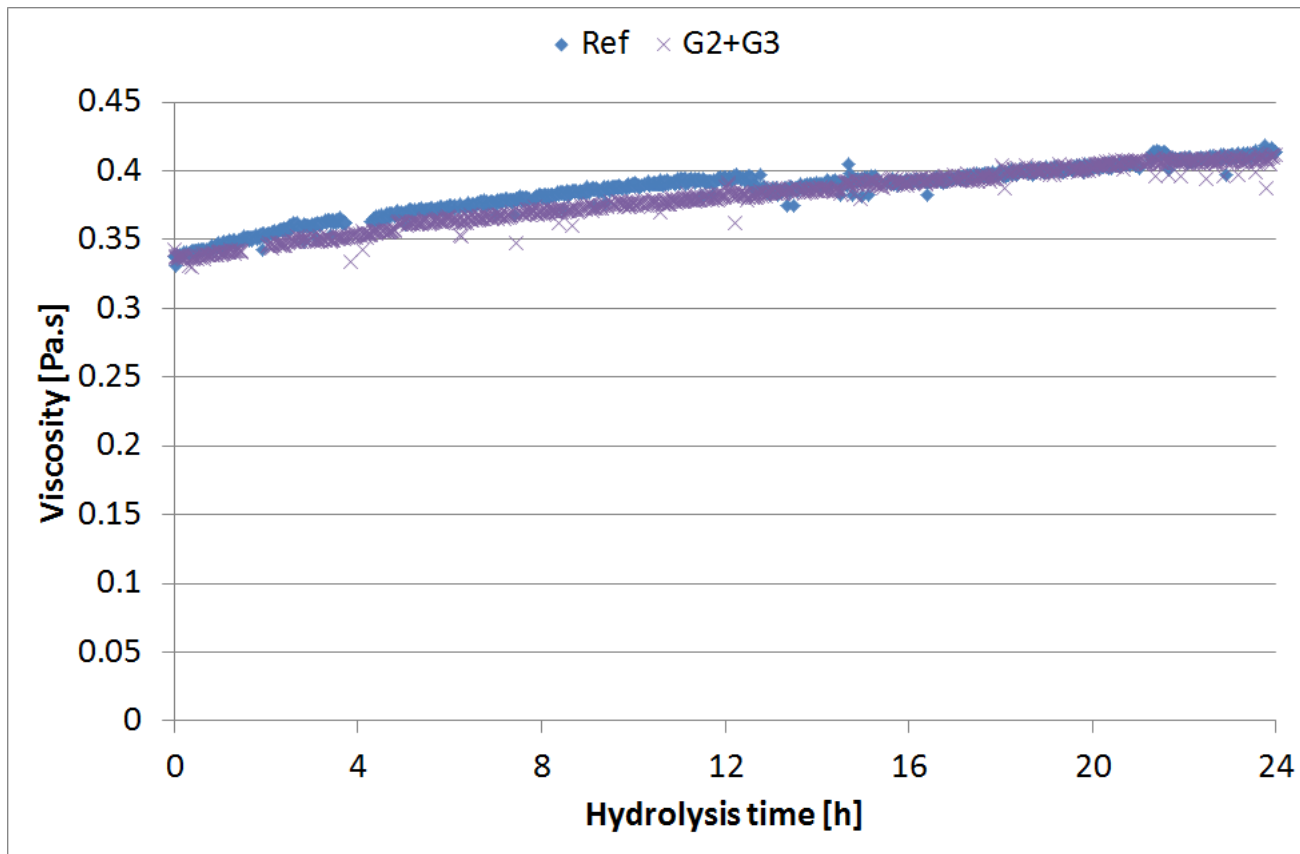


G1 (endo-glucanase) =

Conclusions on filter paper:

- Liquefaction: G2+G3=almost no effect, G1=predominant role.
- G1+G2+G3 no synergy observed
- Ctec2 showed better performance. Hypothesis on enzyme synergist ???

Question: how enzymes act on complex lignocellulosic suspension ?

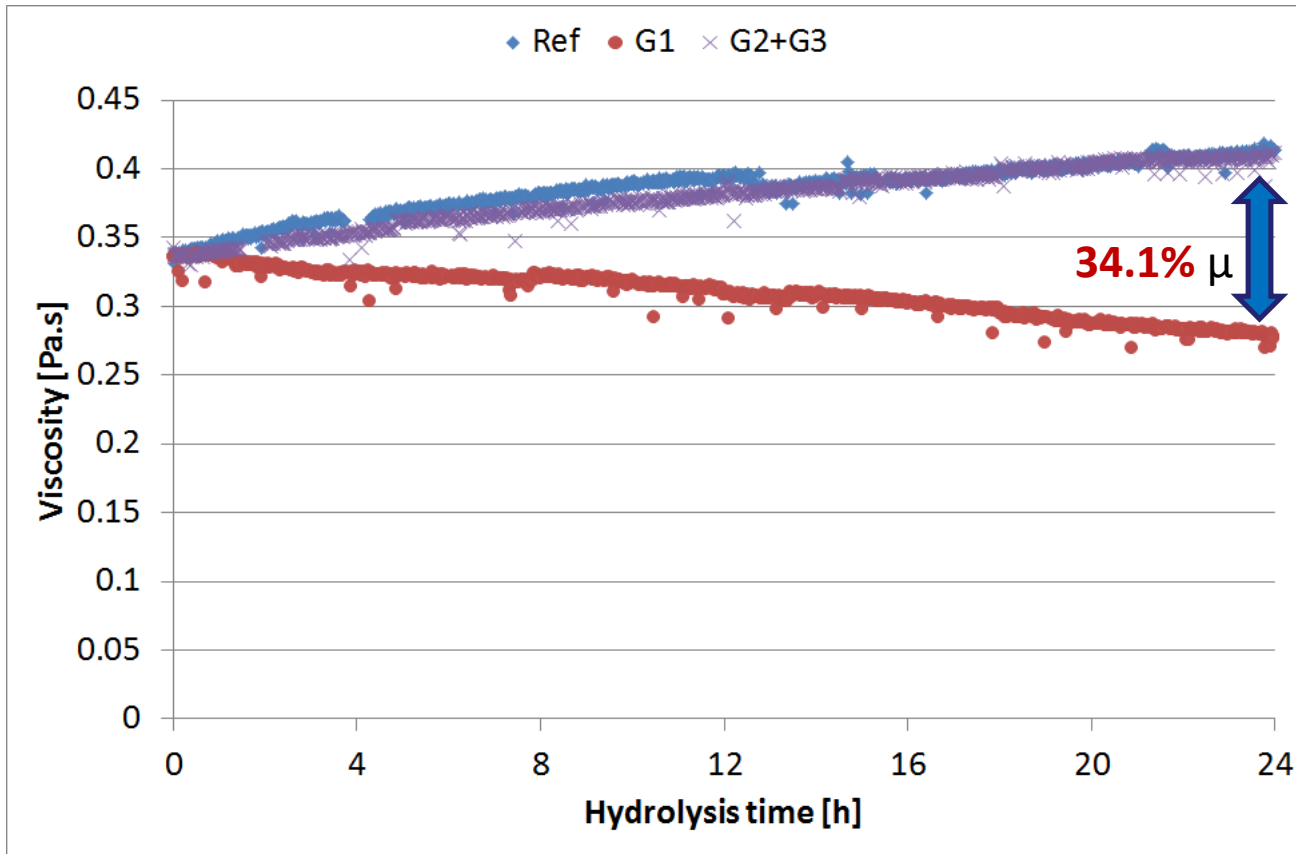


Paper pulp 30gdm/L

Mixing rate 100rpm

Ref: no enzyme

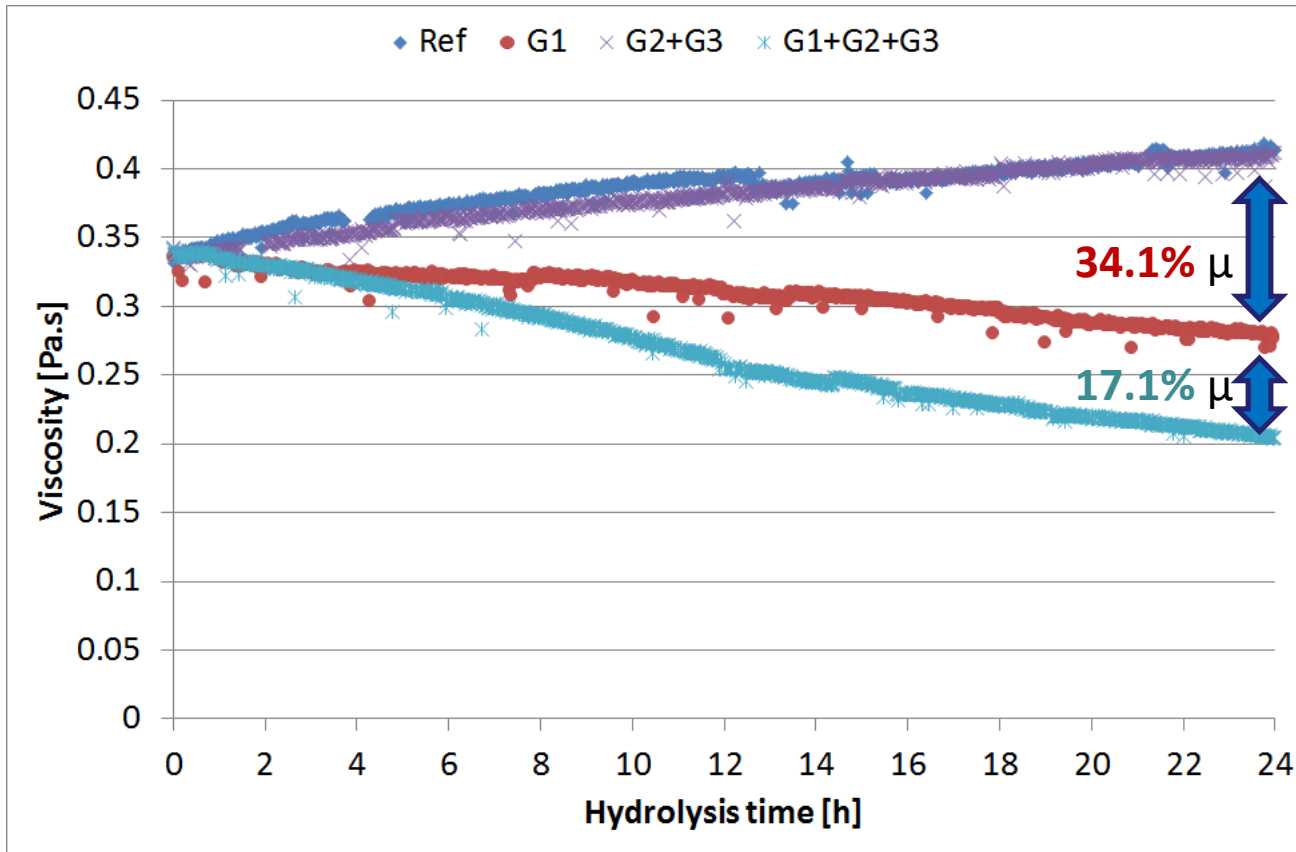
G2+G3 (exo-glucanase & β -glucosidase) = **no contribution.**



Paper pulp 30gdm/L
 Mixing rate 100rpm
 Ref: no enzyme
G2+G3 (exo-glucanase & β -glucosidase) = **no contribution.**



G1 (endo-glucanase) = **34.1%** reduction in μ .



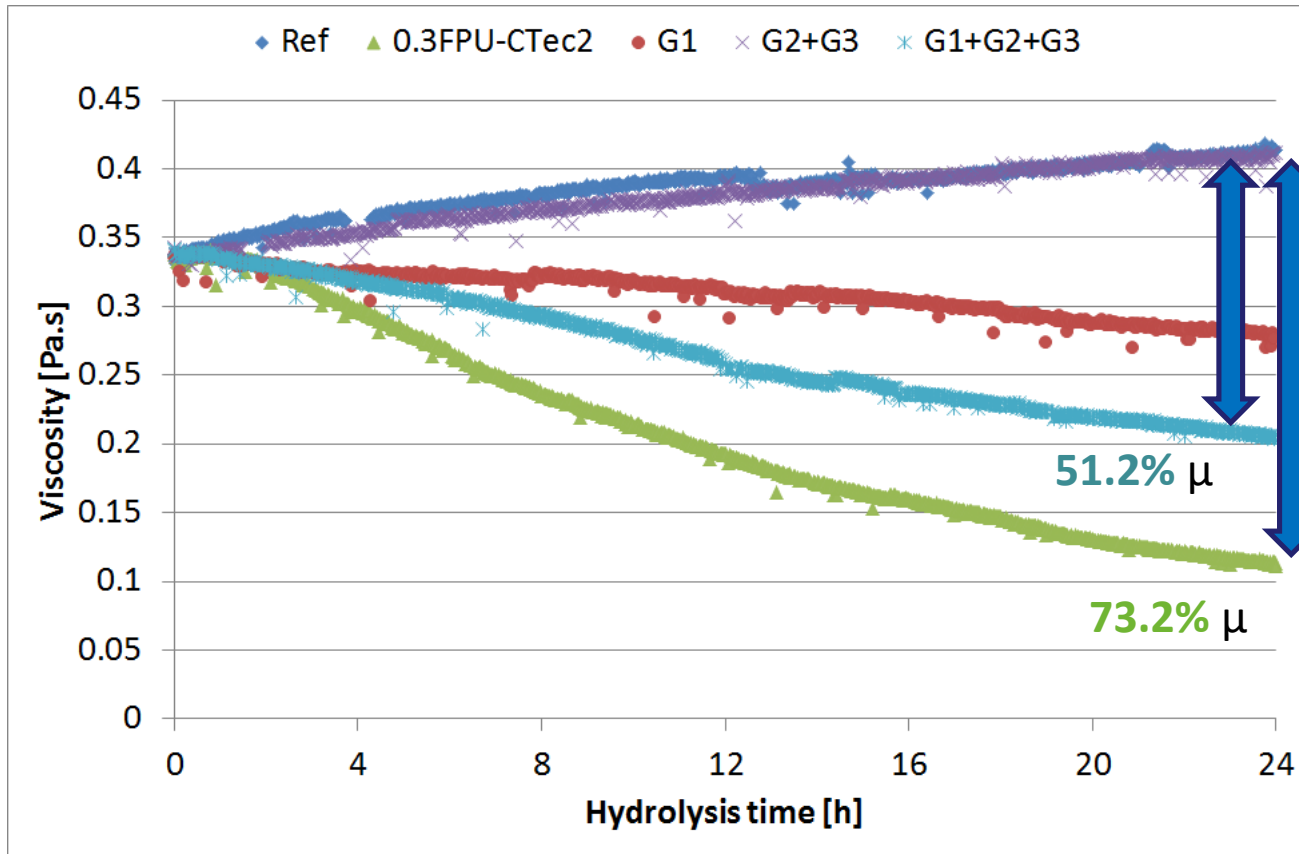
Paper pulp 30gdm/L
Mixing rate 100rpm
Ref: no enzyme
G2+G3 (exo-glucanase & β -glucosidase) = **no contribution.**



G1 (endo-glucanase) = **34.1%** reduction in μ .



Enzyme synergy of **G1+G2+G3**
51.2% reduction in μ = 1.5fold increase vs **G1** alone.

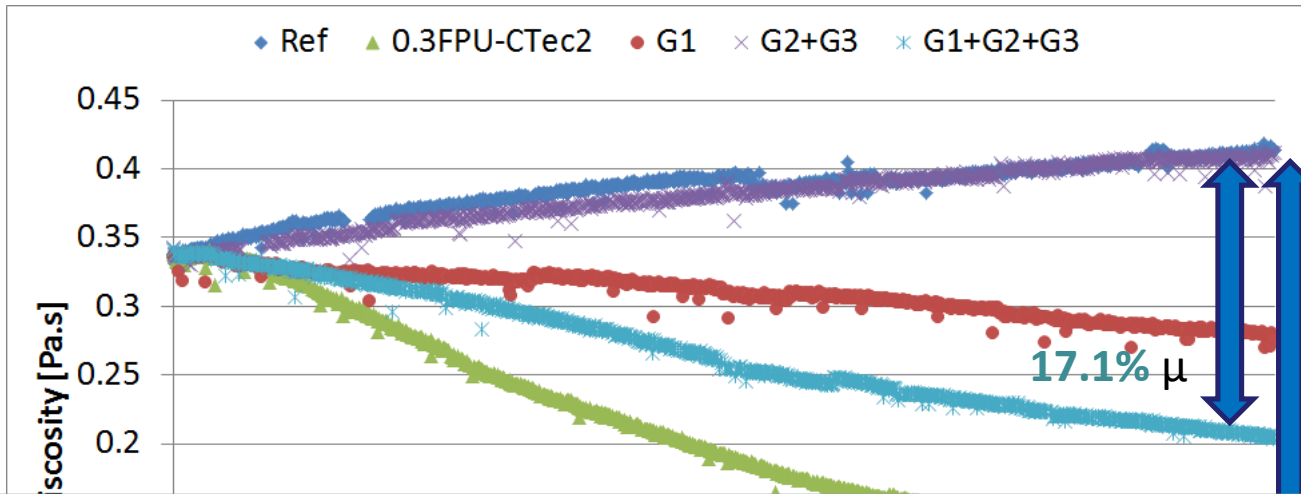


Paper pulp 30gdm/L
Mixing rate 100rpm
Ref: no enzyme
G2+G3 (exo-glucanase & β -glucosidase) = **no contribution**.

G1 (endo-glucanase) = **34.1%** reduction in μ .

Enzyme synergy of **G1+G2+G3**
51.2% reduction in μ = 1.5fold increase vs **G1** alone.

Ctec2 >> **G1+G2+G3**
73.2% reduction in μ .



Paper pulp 30gdm/L

Mixing rate 100rpm

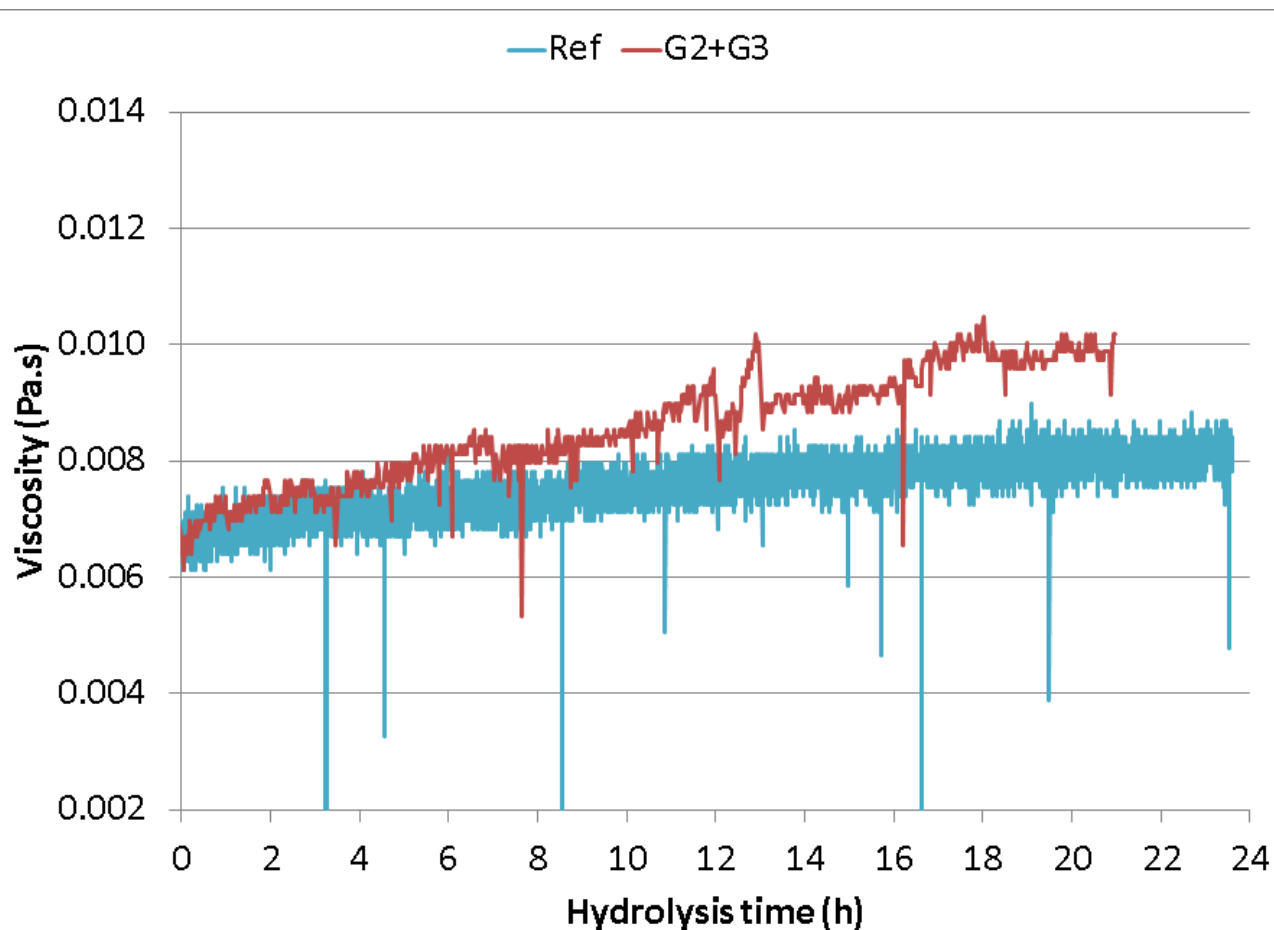
Ref: no enzyme

G2+G3 (exo-glucanase & β -glucosidase) = **no contribution.**

G1 (endo-glucanase) = **20.6%** reduction in μ .

Conclusions paper pulp

1. **G2+G3 = no effect, G1 = predominant**
2. **Synergy observed on G1+G2+G3.**
3. **Ctec2 = stronger liquefaction efficiency. It is likely explained by higher activities content of exo-glucanase and β -glucosidase.**

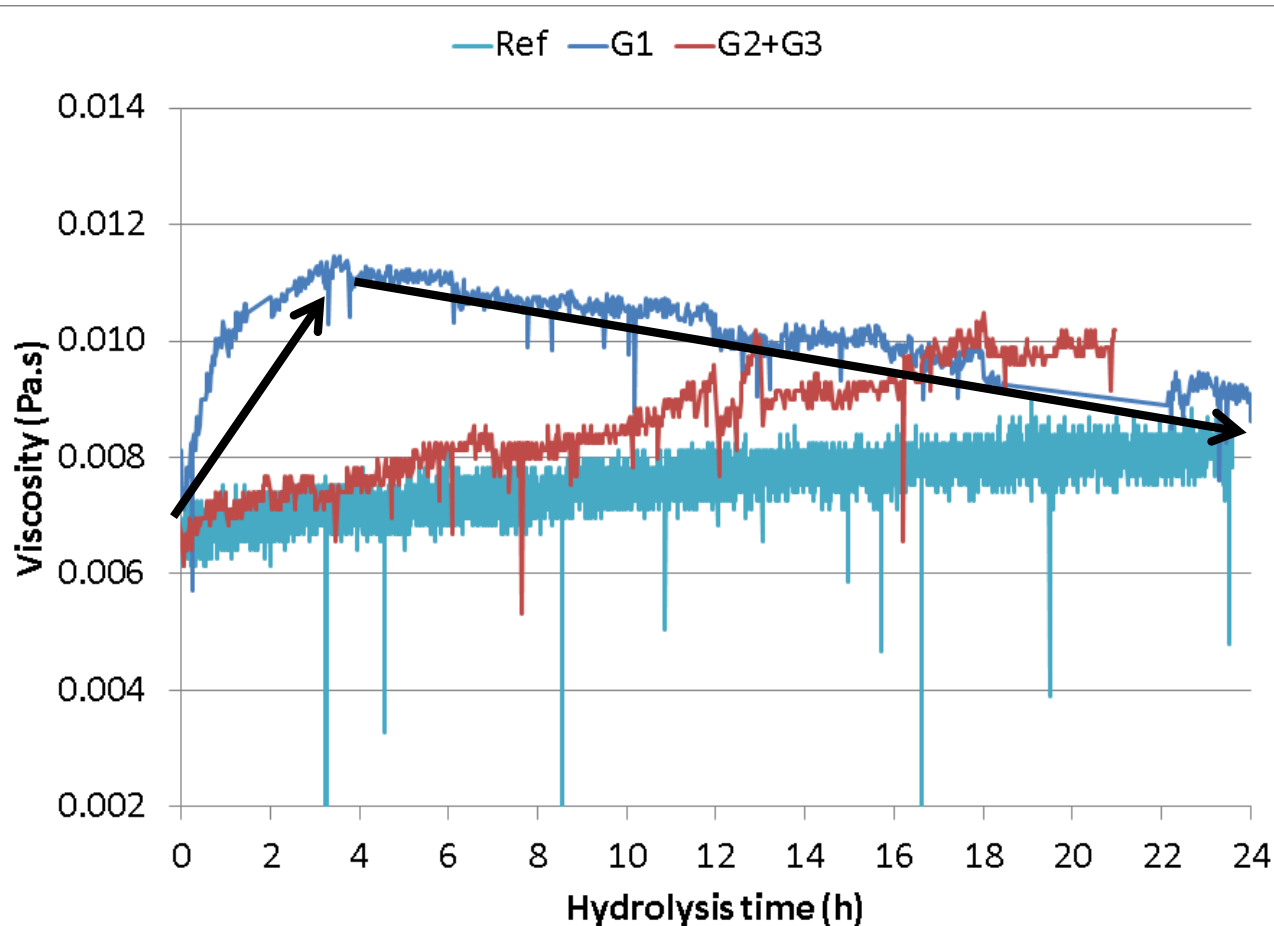


Sugarcane bagasse 30gdm/L

Mixing rate 100rpm

Ref: no enzyme

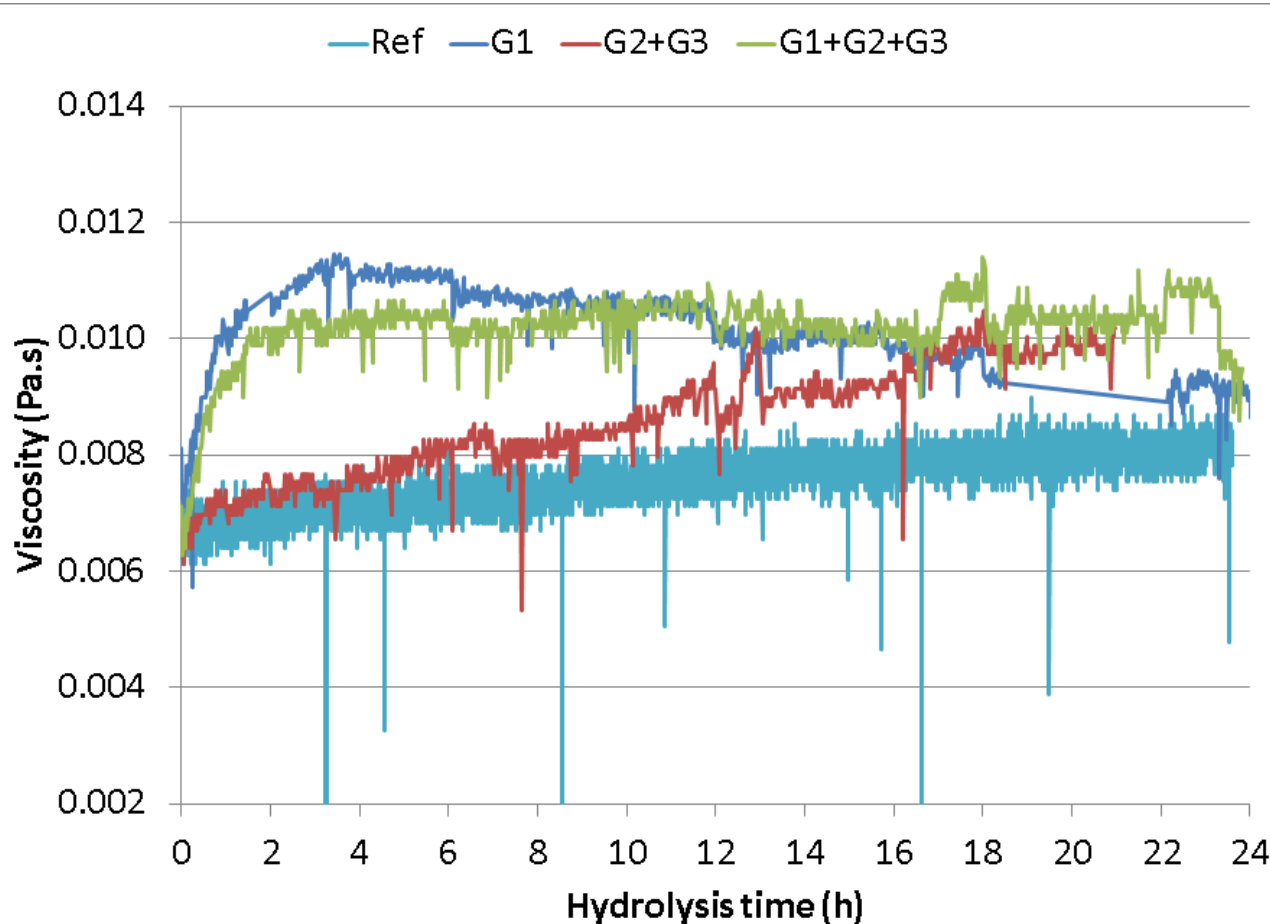
G2+G3 (exo-glucanase & β -glucosidase) = **gradual rise in viscosity.**



Sugarcane bagasse 30gdm/L
Mixing rate 100rpm
 Ref: no enzyme
G2+G3 (exo-glucanase & β -glucosidase) = **small contribution.**



G1 (endo-glucanase) = **rise** of suspension viscosity in the first 3h.



Sugarcane bagasse 30gdm/L

Mixing rate 100rpm

Ref: no enzyme

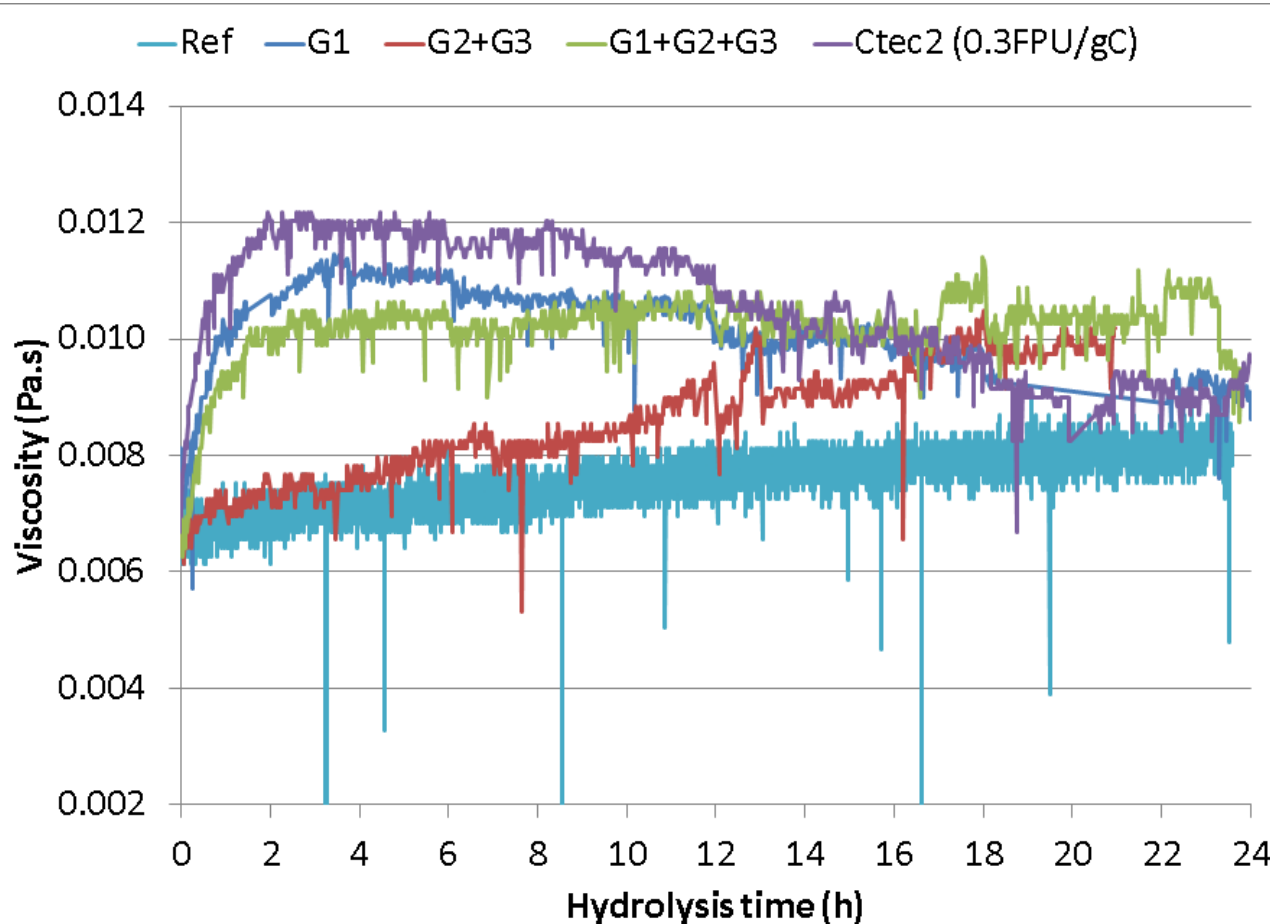
G2+G3 (exo-glucanase & β -glucosidase) = **small contribution.**



G1 (endo-glucanase) = **rise** of suspension viscosity in the first 3h.



G1+G2+G3 similar viscosity pattern with either **G1** alone or **Ctec2**.



Sugarcane bagasse 30gdm/L

Mixing rate 100rpm

Ref: no enzyme

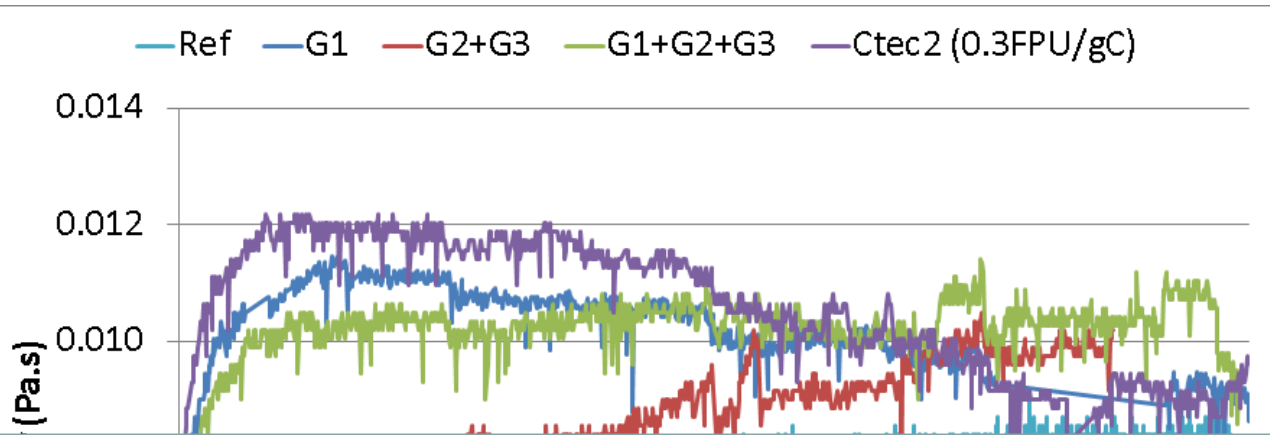
G2+G3 (exo-glucanase & β -glucosidase) = **small contribution.**



G1 (endo-glucanase) = **rise** of suspension viscosity in the first 3h.



G1+G2+G3 similar viscosity pattern with either **G1** alone or **Ctec2**.

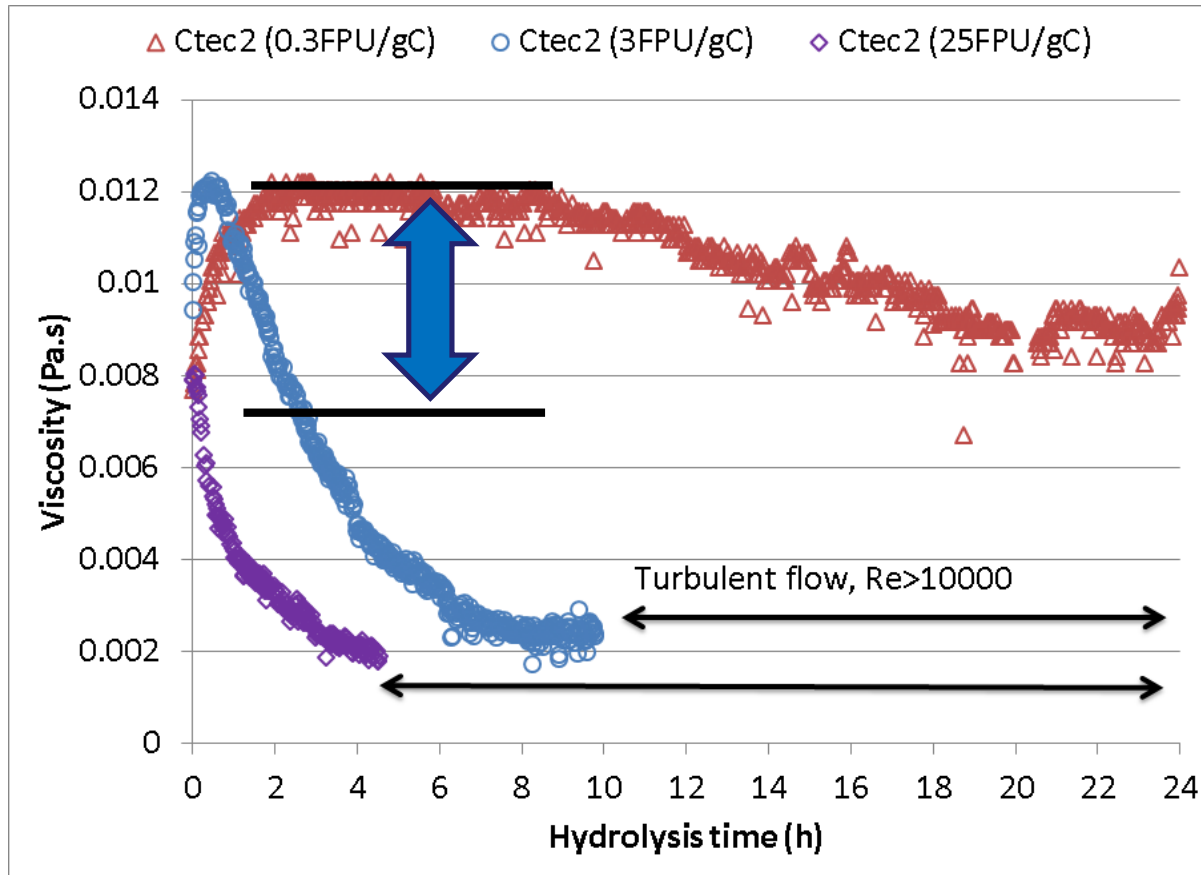


Sugarcane bagasse 30gdm/L
Mixing rate 100rpm
Ref: no enzyme
G2+G3 (exo-glucanase & β -glucosidase) = **small contribution.**



Message 2:

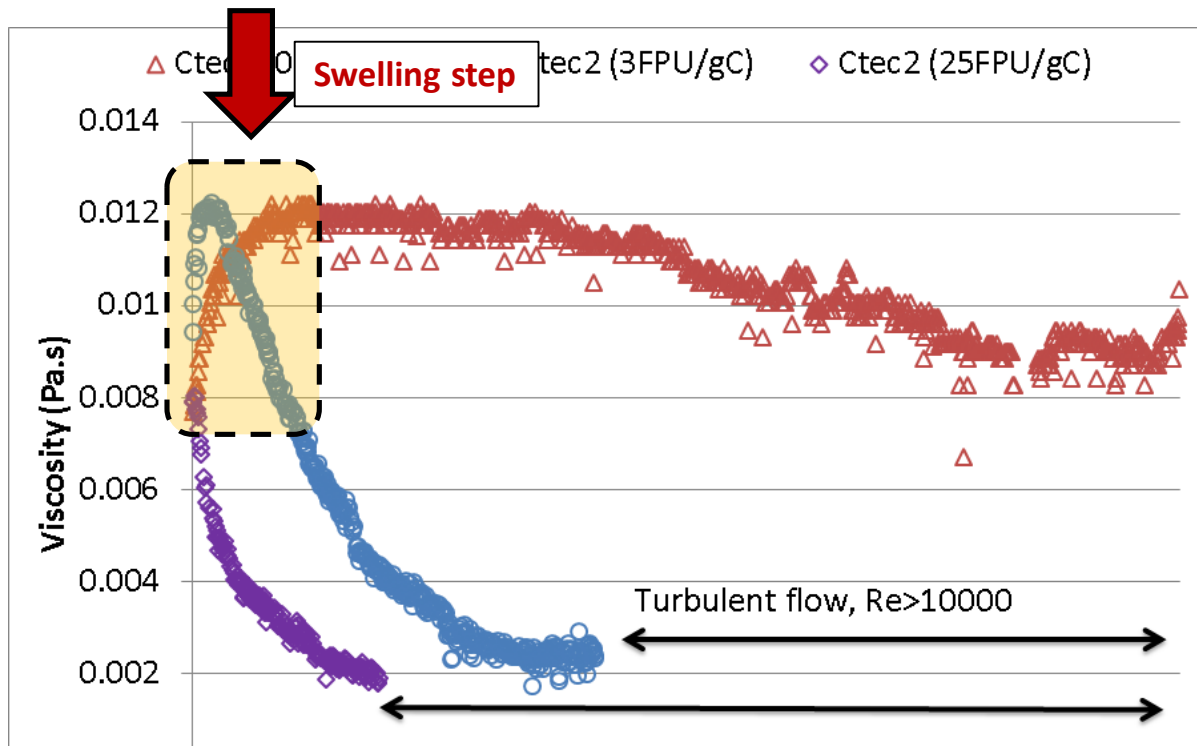
- Expected results :
 - G1=predominant role, G2+G3=minor contribution.
 - Synergy between G1, G2 and G3 → liquefaction efficiency.
- Original insight:
 - Swelling step of sugarcane bagasse. Mechanism???



SCB 30gdm/L
Mixing rate 100rpm

No rise in viscosity
observed at high enzyme
loading (25FPU/gC)

Same level of increase in
viscosity with 0.3 & 3
FPU/gC



SCB 30gdm/L
Mixing rate 100rpm

No rise in viscosity
observed at high enzyme
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Same level of increase in
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FPU/gC

From literature:

H1: \uparrow [s] lead to \uparrow viscosity (Fan et al., 2003; T.C. Nguyen et al. 2013, this work)

H2: \uparrow particle size lead to \uparrow viscosity (Dasari and Berson 2007)

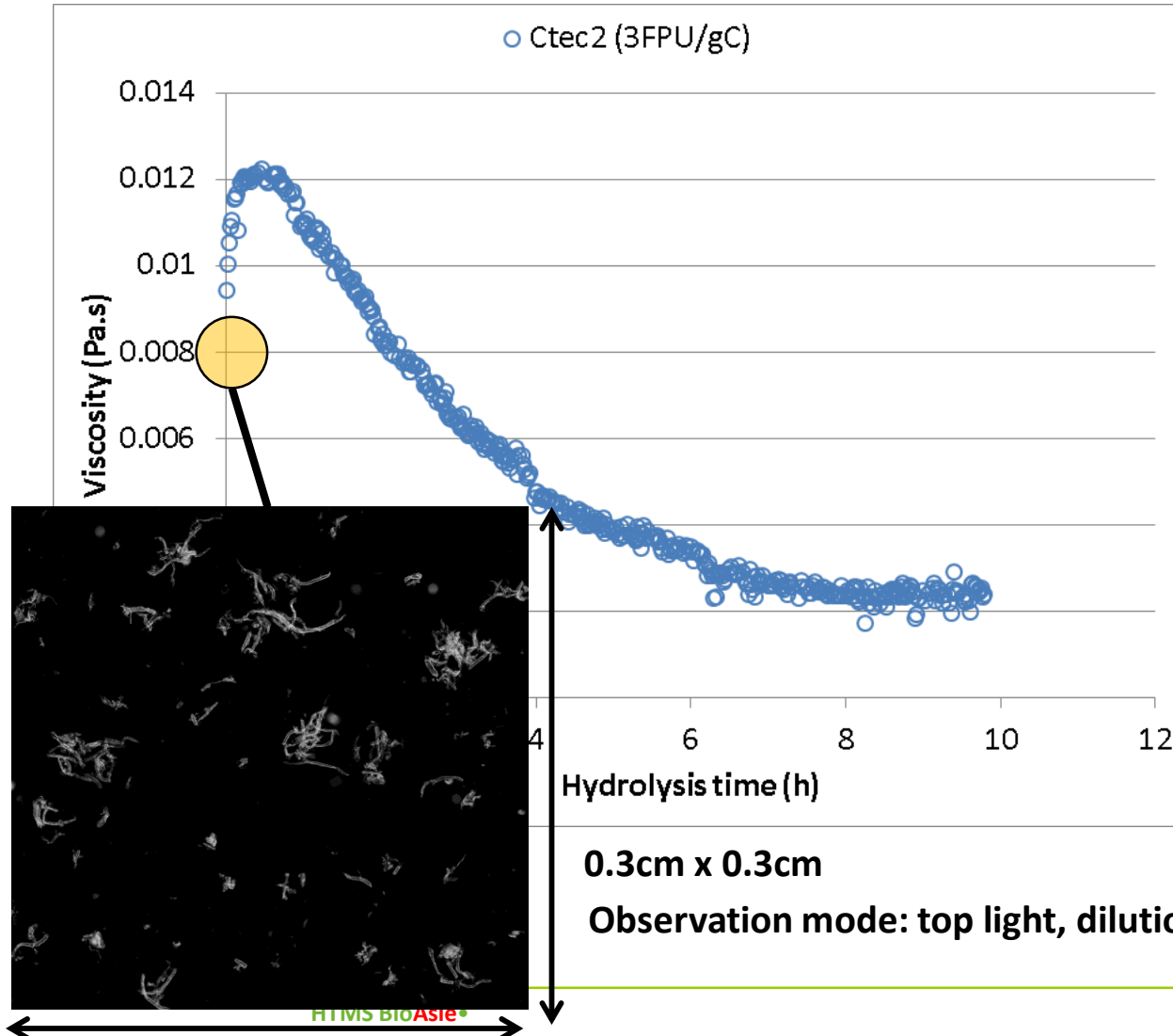


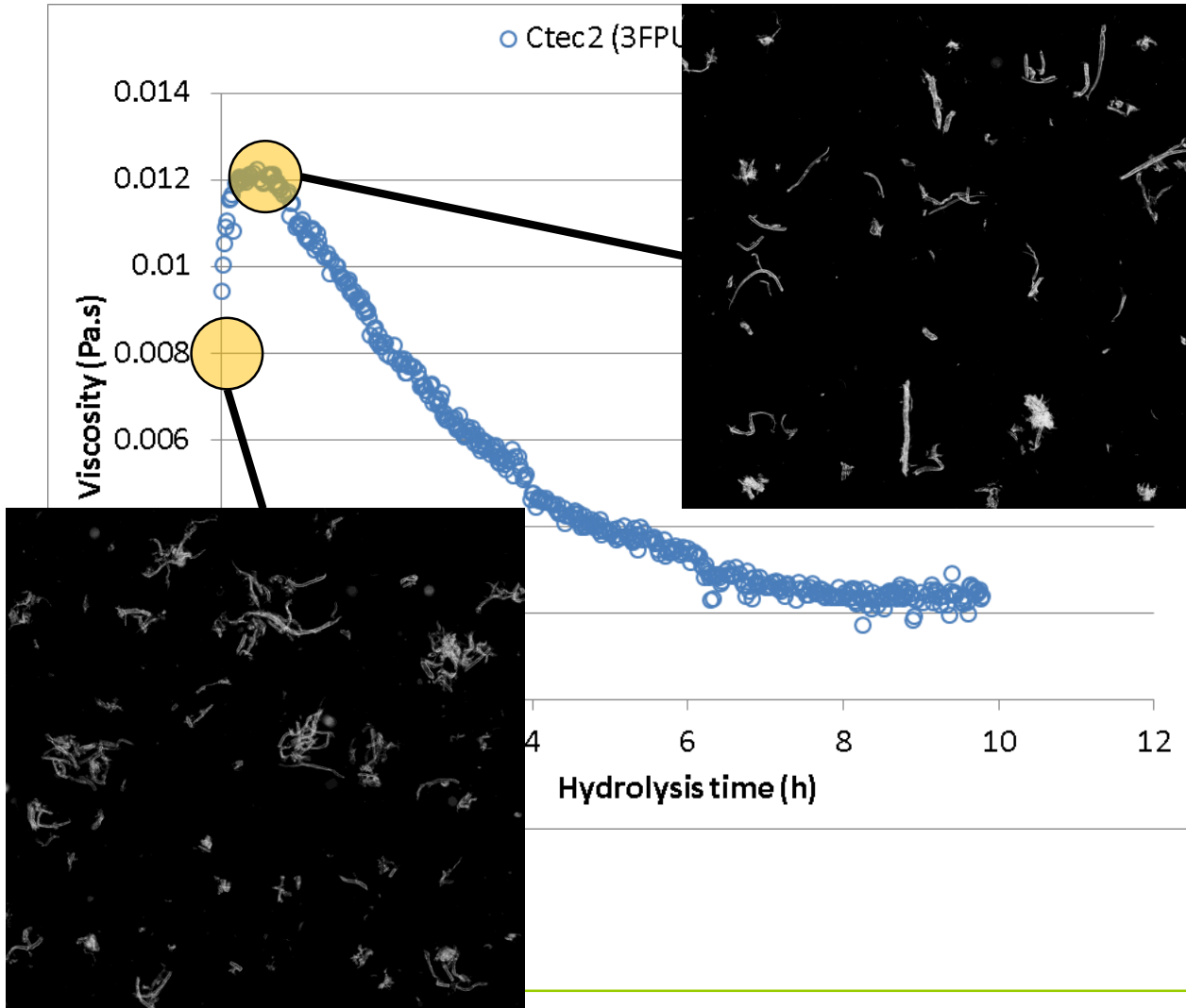
Need to be verified

Data from Morphology

At 0h:

- Considerable number of agglomerates.
- Mean dCE= $12.98 \pm 1.34 \mu\text{m}$





Data from Morphology

At 0h:

- Considerable number of agglomerates.
- Mean dCE= $12.98 \pm 1.34 \mu\text{m}$

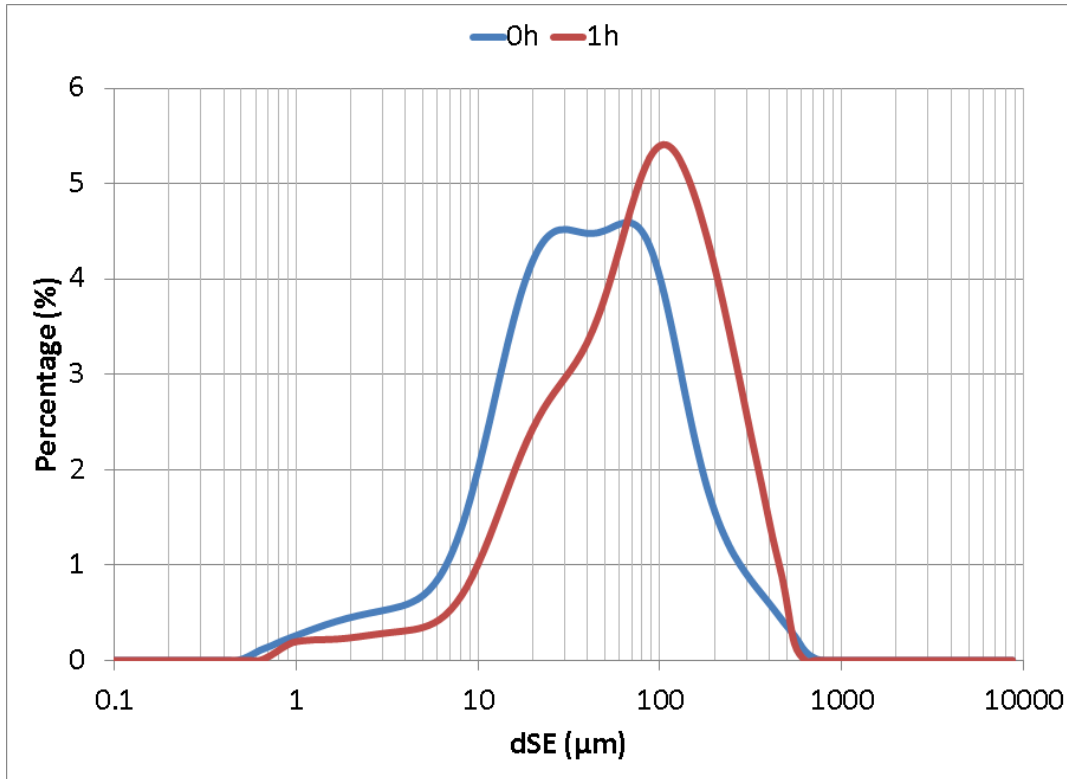
At 1h:

- Few agglomerates, several individuals particles.
- Mean dCE= $18.76 \mu\text{m}$



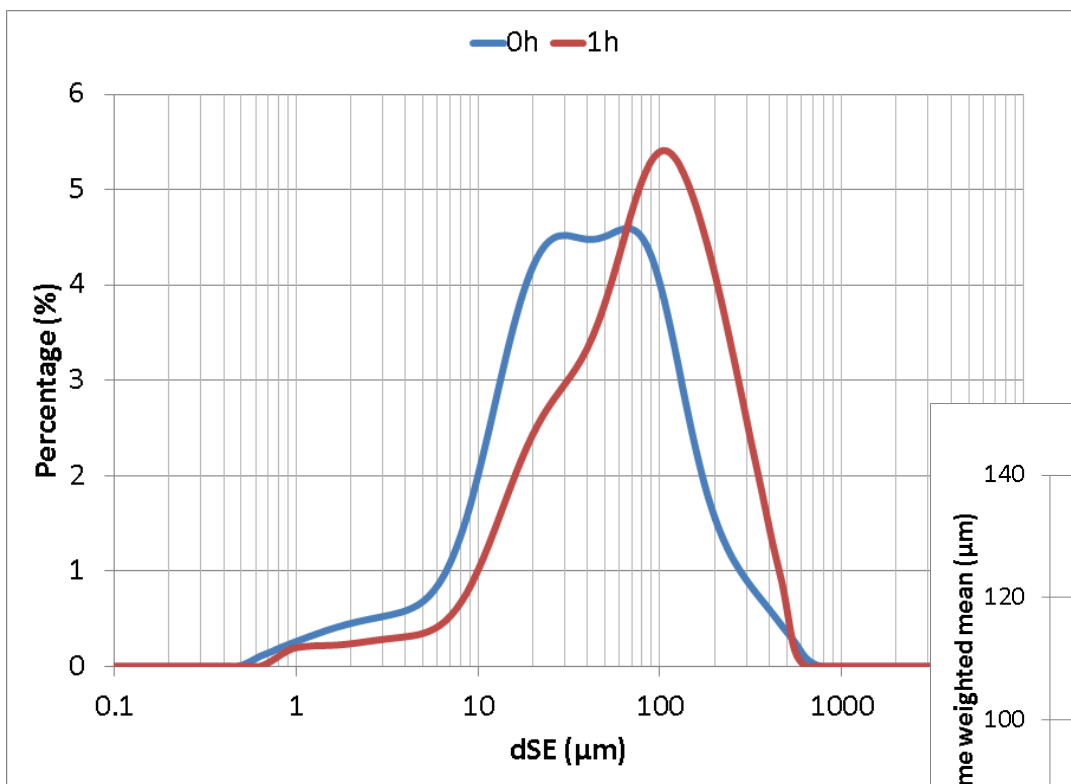
Increase in mean dCE
Disappear of agglomerates

Data from DLS

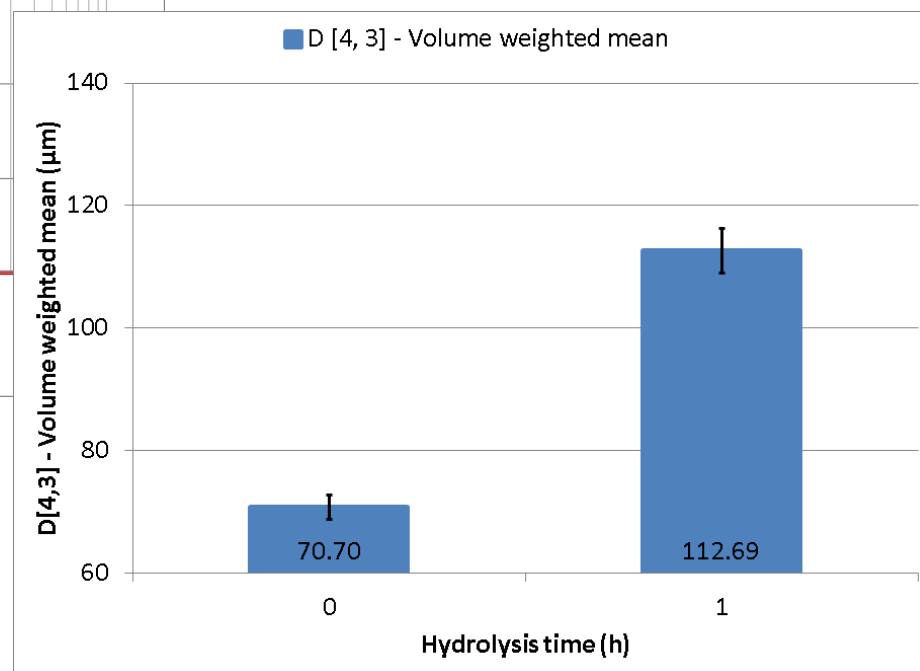


dSE = diameter of sphere equivalent.

Data from DLS

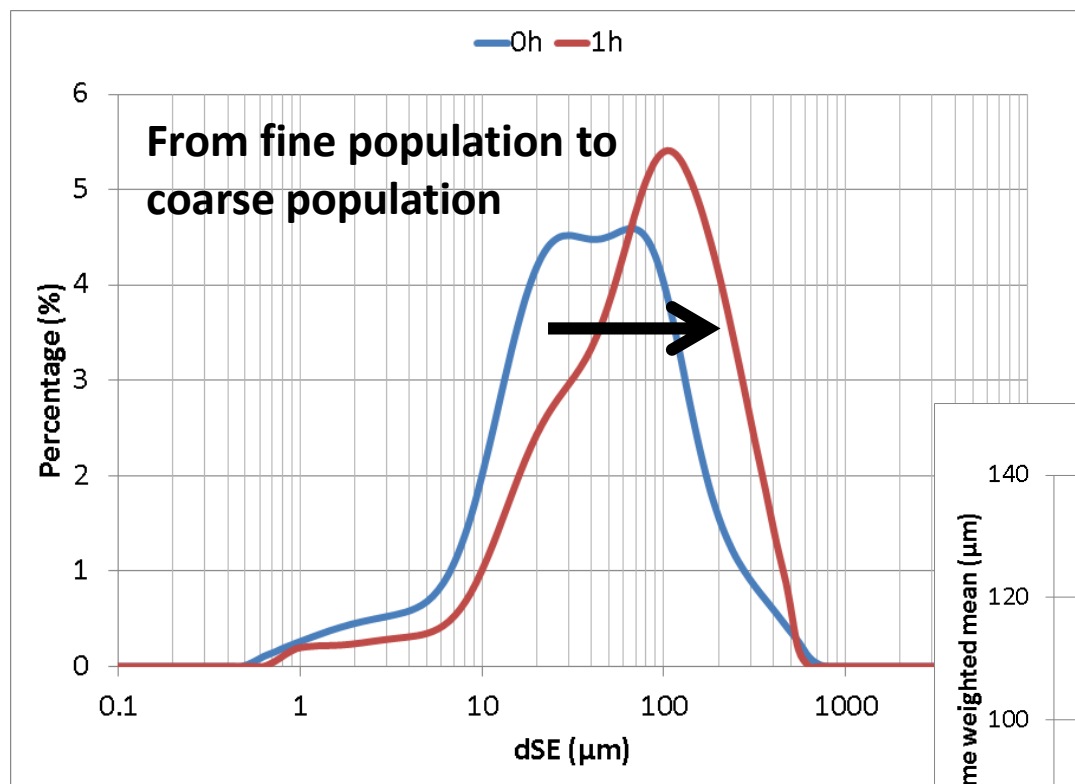


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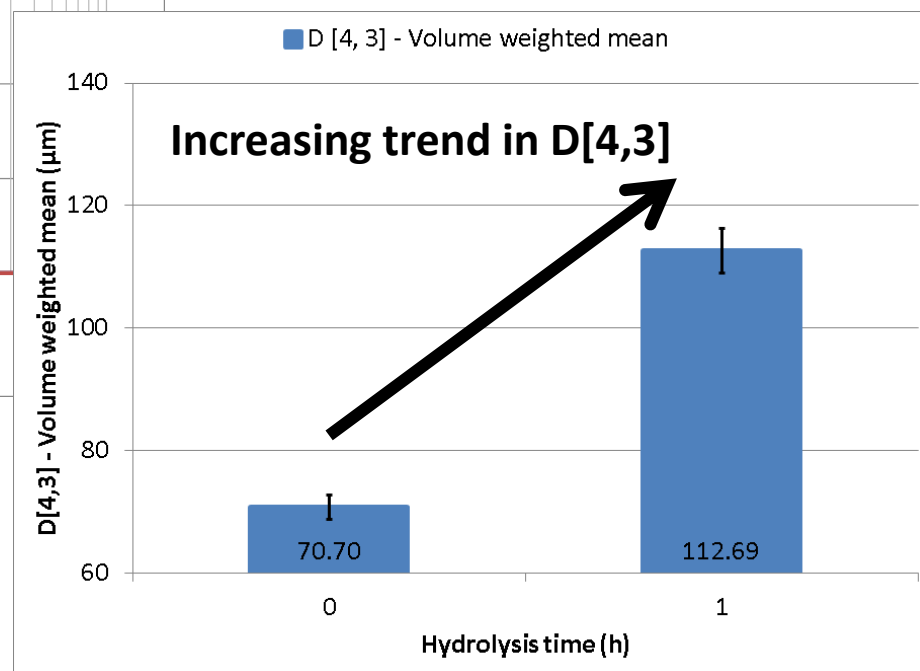


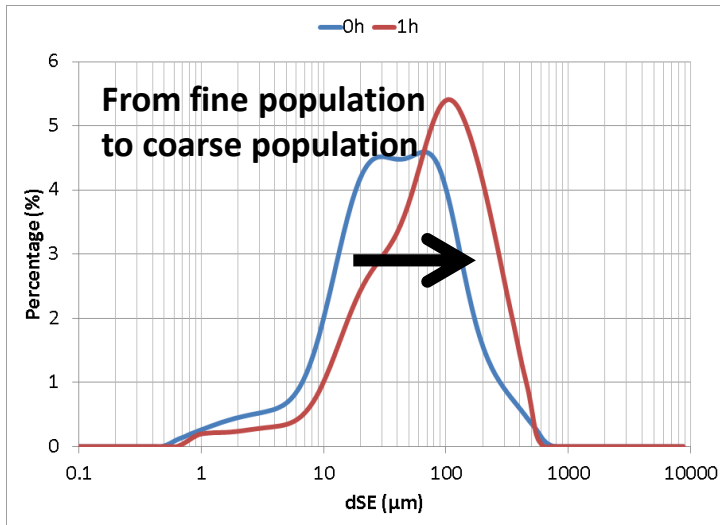
Data from DLS

Solubilization of fine population.



dSE = diameter of sphere equivalent.





Phenomenon observed: rise in viscosity

Hypothesis 2

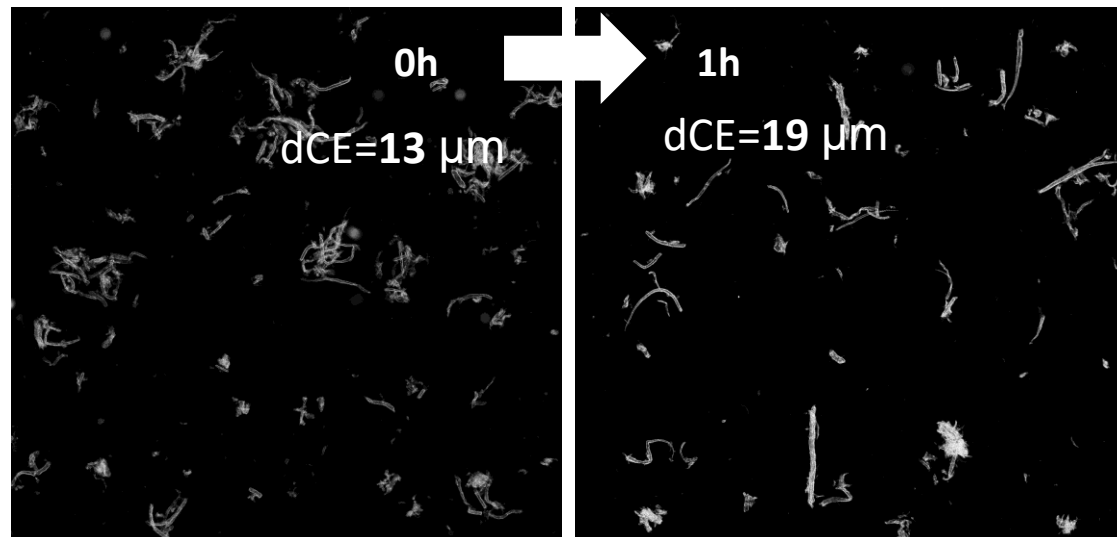
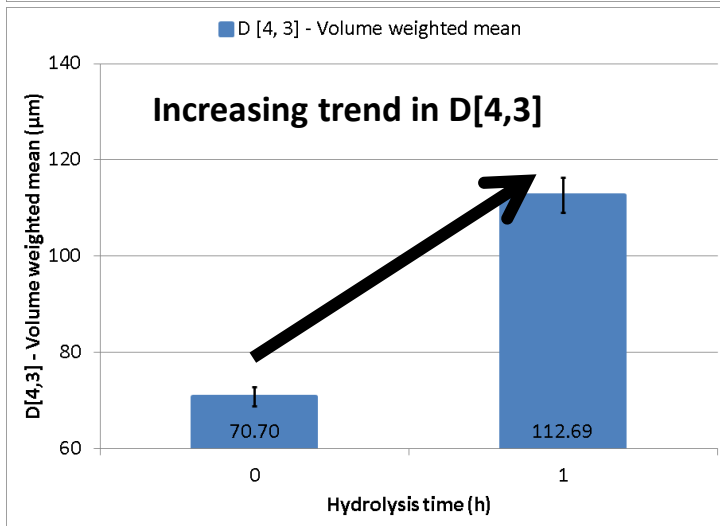


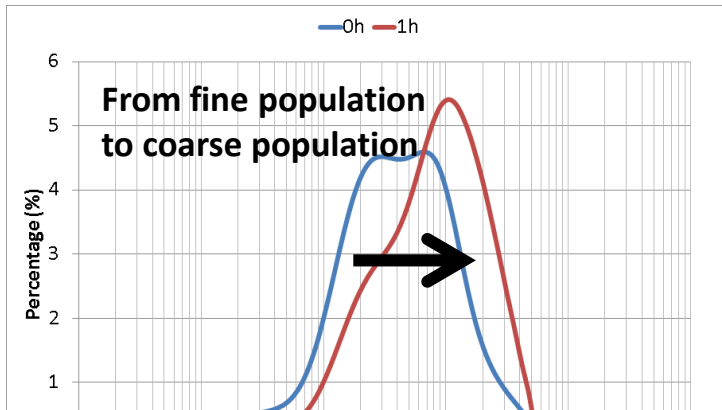
Analysis results

Cellulose to glucose conversion $3.45 \pm 0.45\%$

→ Negligible variation of **[S]**

DLS + Morpho: increase in particle size

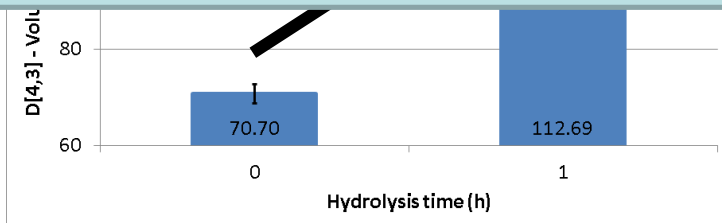




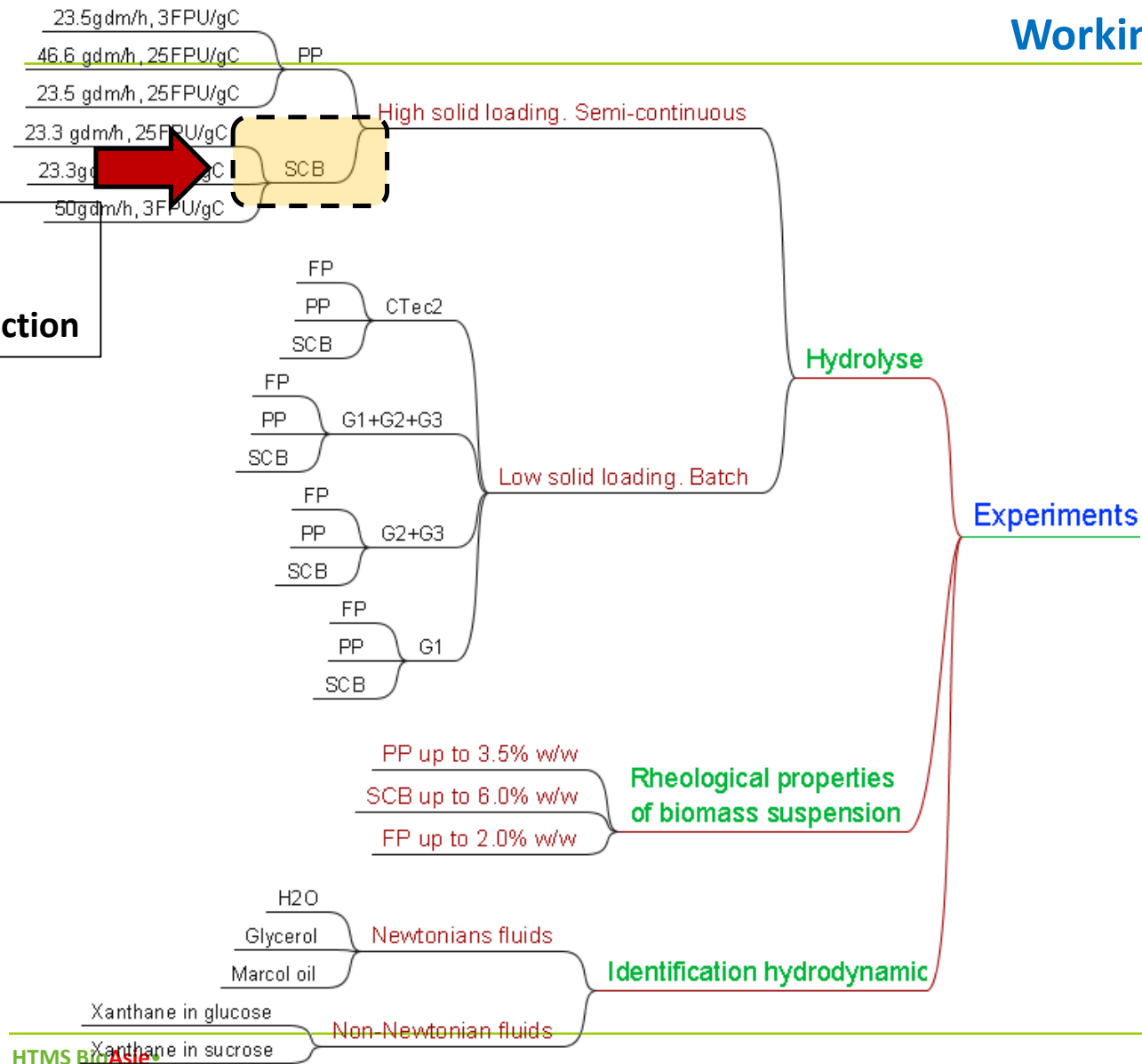
- Solubilization of fine population.
- Increase in particle mean diameter
- Particles were separated from agglomerates

Message 3

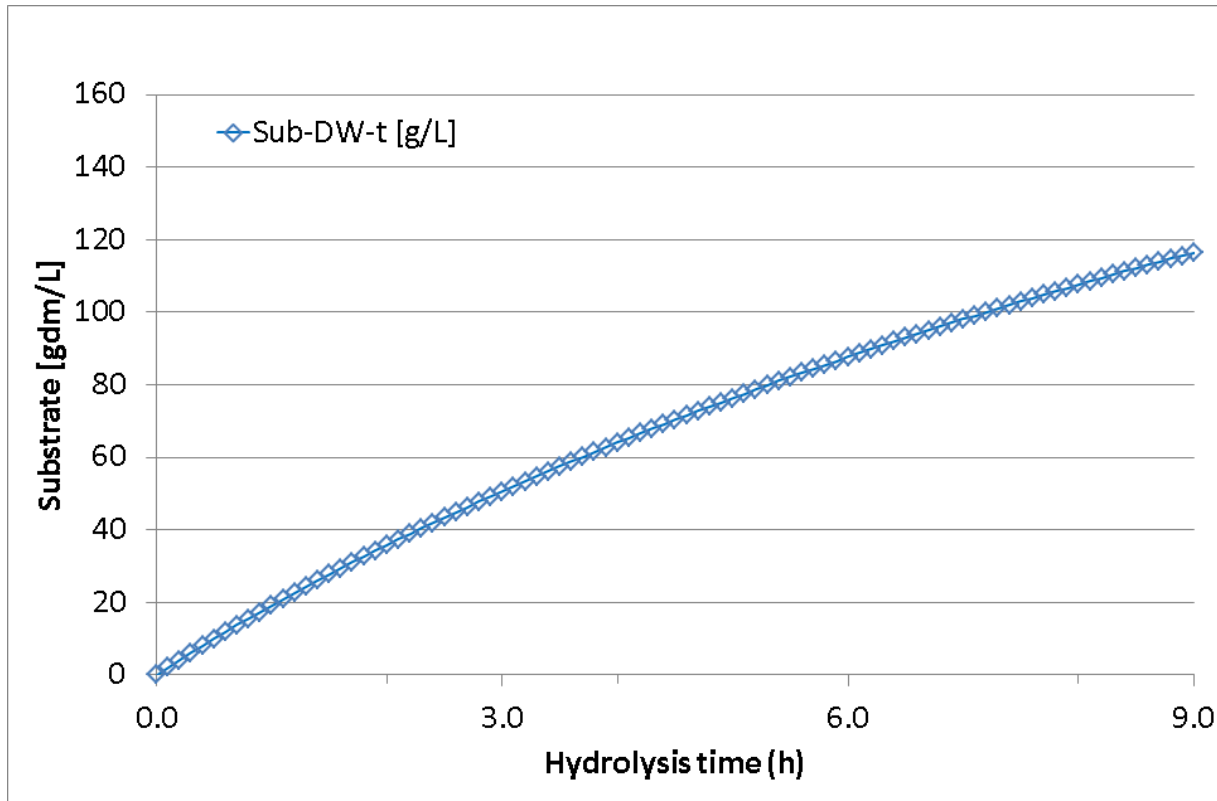
- Swelling step on SCB suspension = evolution of population from fine to coarse
- Obtained results are not enough to reveal its mechanism.



Message 4:
Mechanism of
enzymatic liquefaction

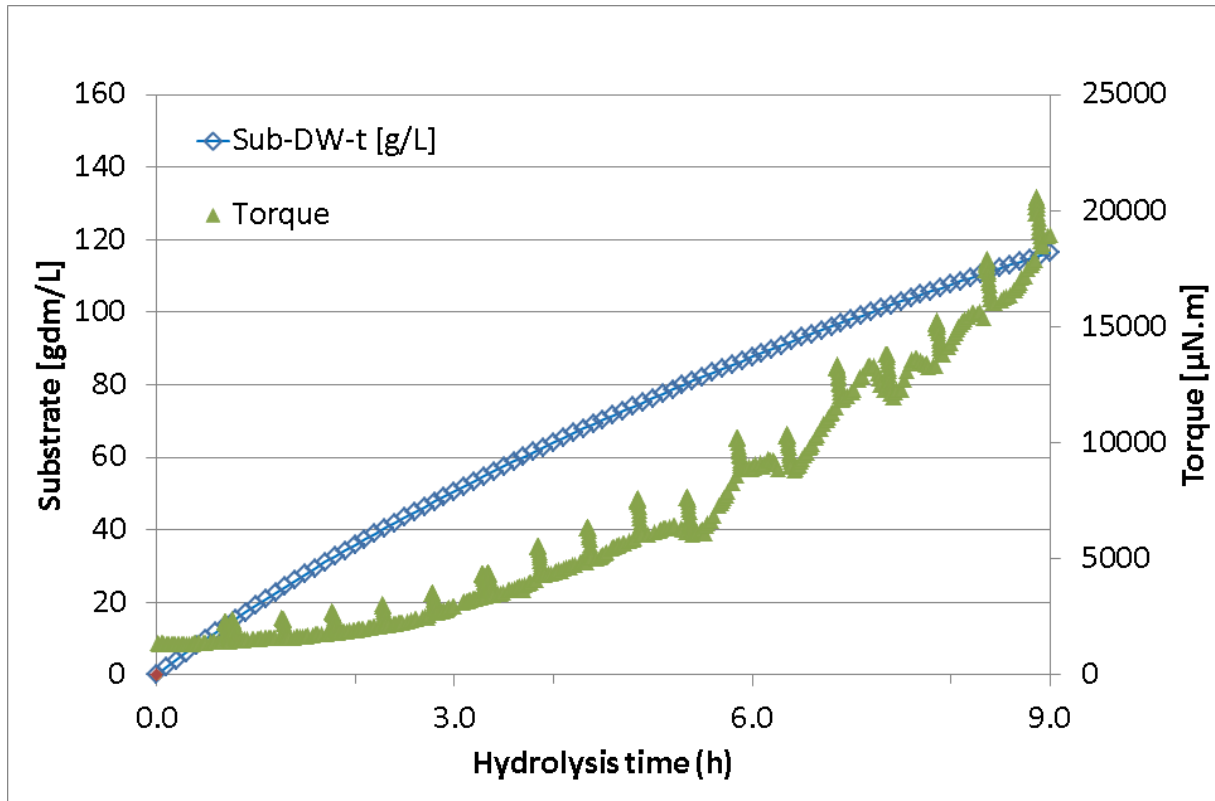


Experiment overview: from raw to interpreted data



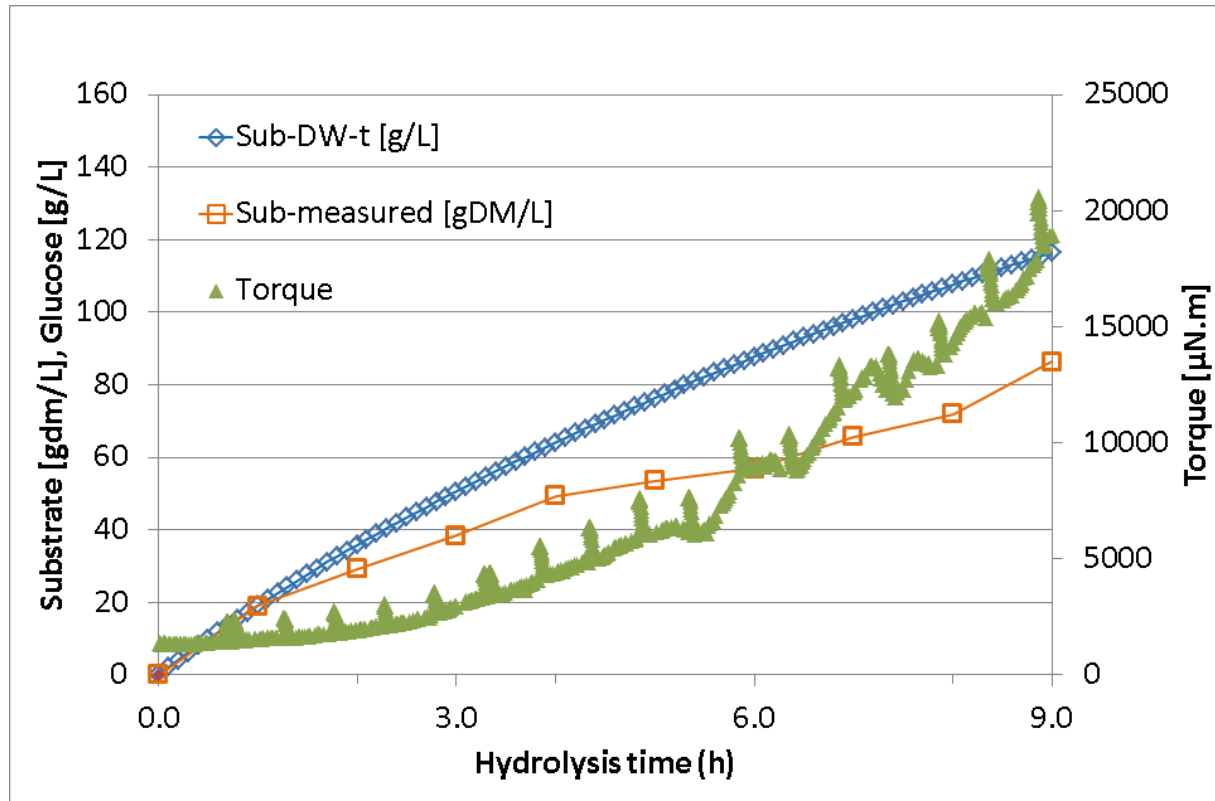
- **Substrate** was feed at constant rate Q_s (g/h)
- **Enzyme** was feed with substrate at fixed ratio $Q_e/Q_s = \text{const}$

Experiment overview: from raw to interpreted data



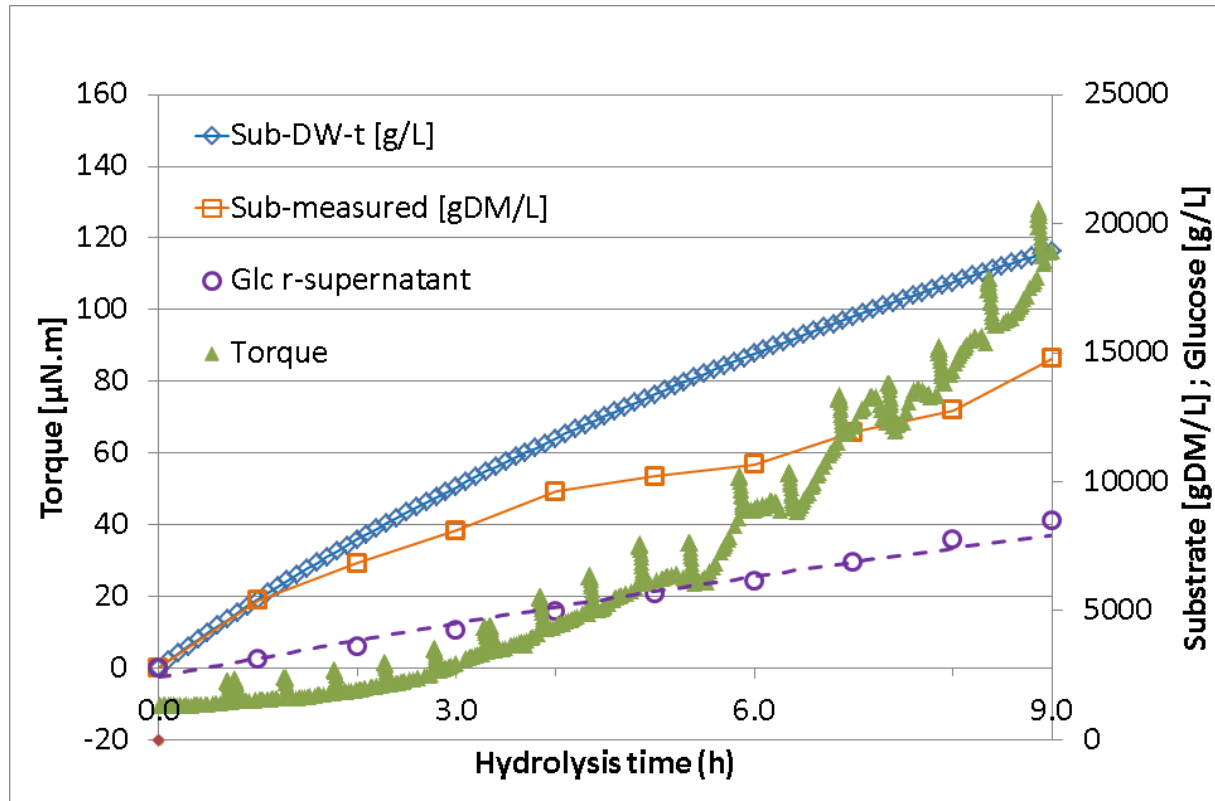
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- **Torque** values were recorded every 1min

Experiment overview: from raw to interpreted data



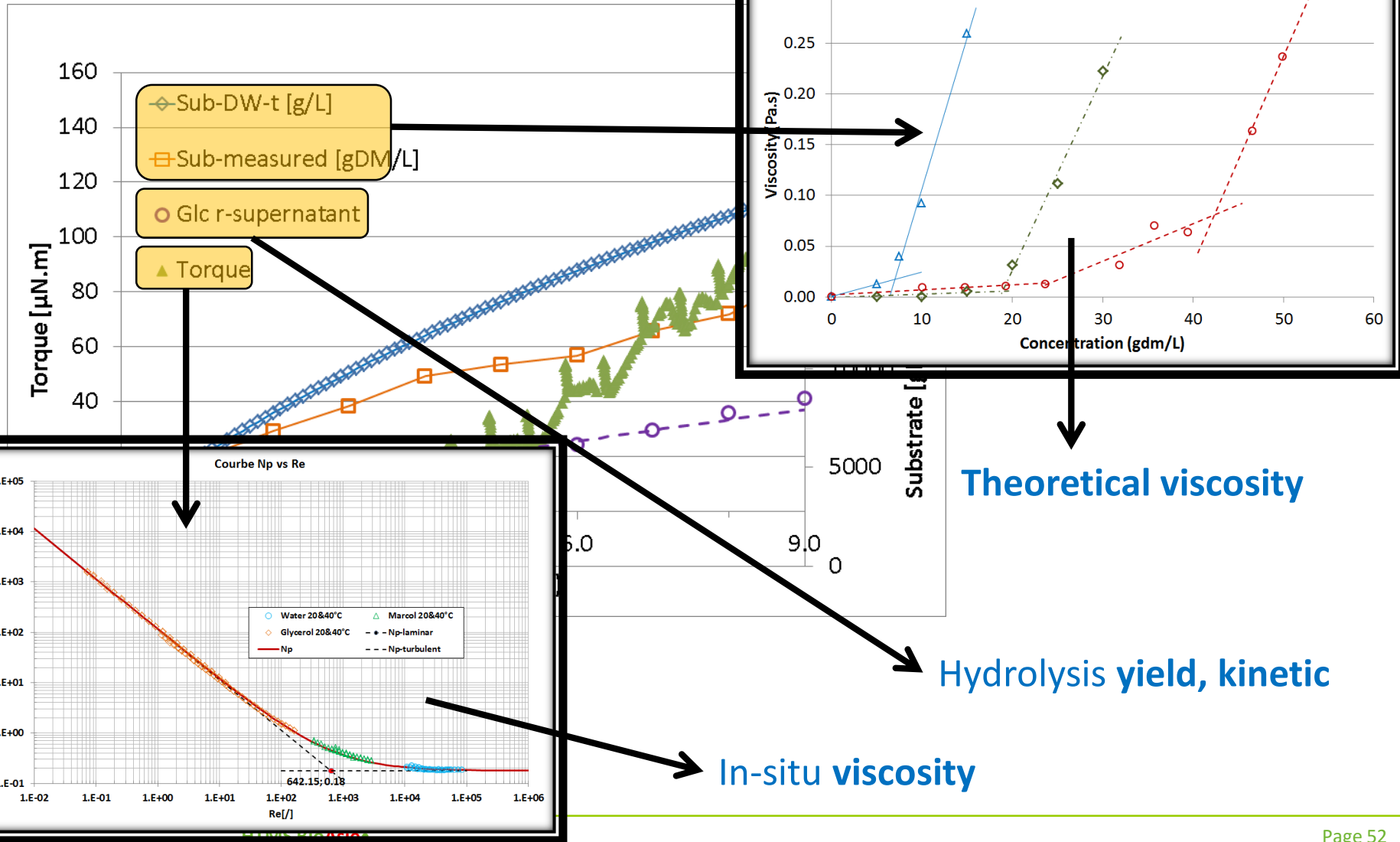
- **Substrate** was feed at constant rate Q_s (g/h)
- **Enzyme** was feed with substrate at fixed ratio $Q_e/Q_s = \text{const}$
- **Torque** values were recorded every 1min
- Periodically, samples were taken for analysis of **dry matter content**

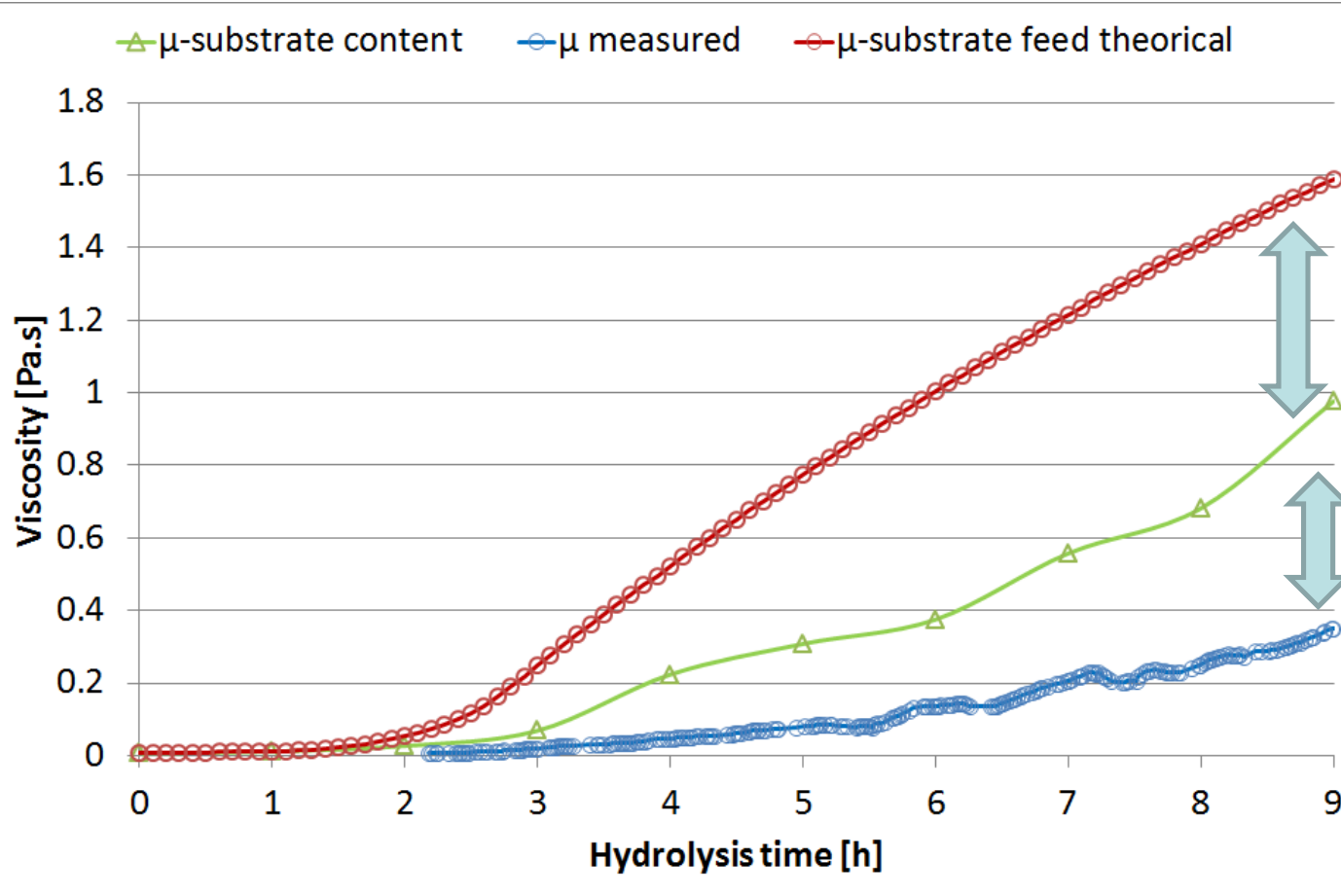
Experiment overview: from raw to interpreted data



- **Substrate** was feed at constant rate Q_s (g/h)
- **Enzyme** was feed with substrate at fixed ratio $Q_e/Q_s = \text{const}$
- **Torque** values were recorded every 1min
- Periodically, samples were taken for analysis of **dry matter content** and **glucose**

Experiment 1

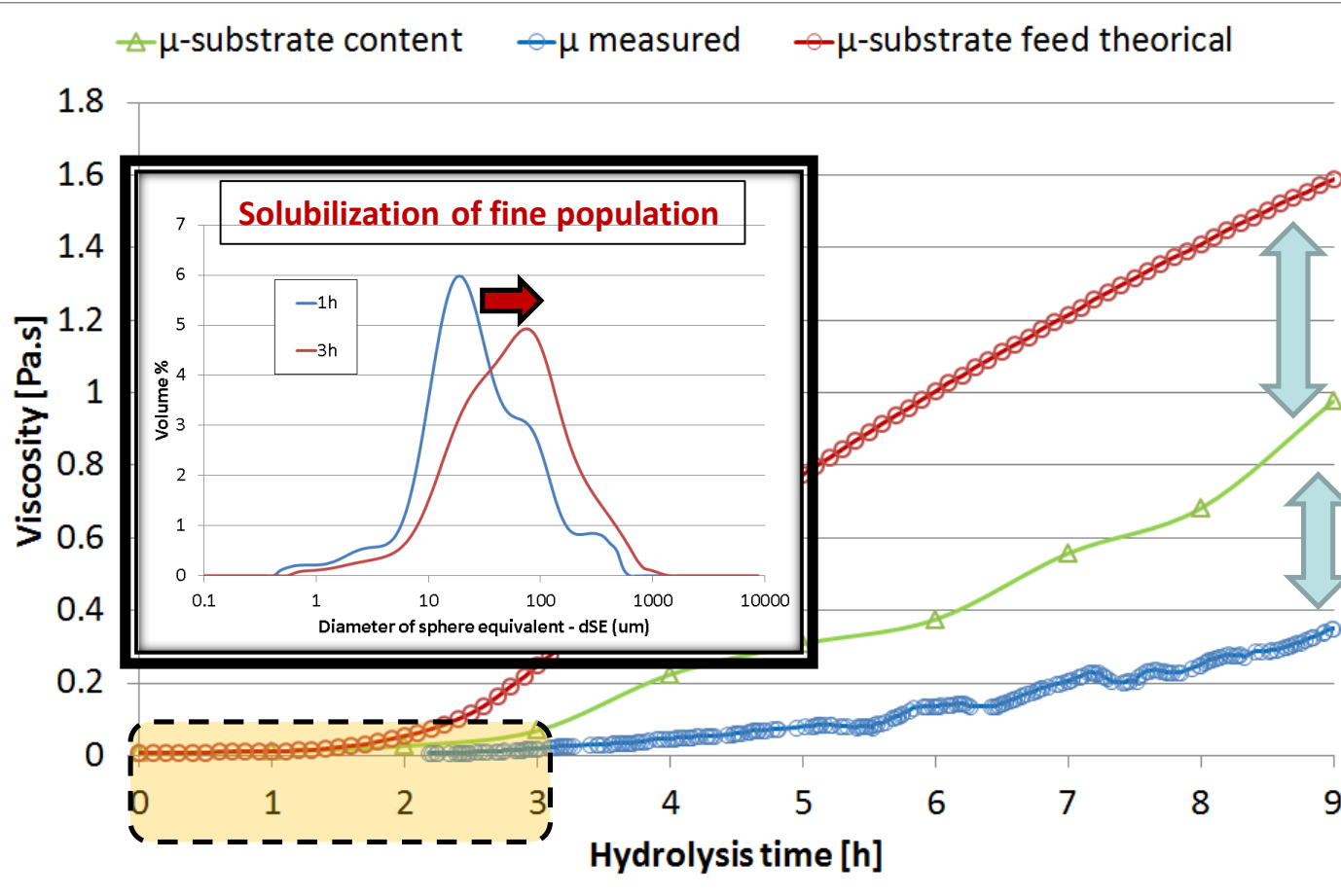




Final [SCB] **140** gdm/L
 Qs = **23.3** gdm/h
 Enzyme: **25**FPU/g cellulose
Phase feeding

Substrate solubilization

Fiber modification



Final [SCB] **140** gdm/L

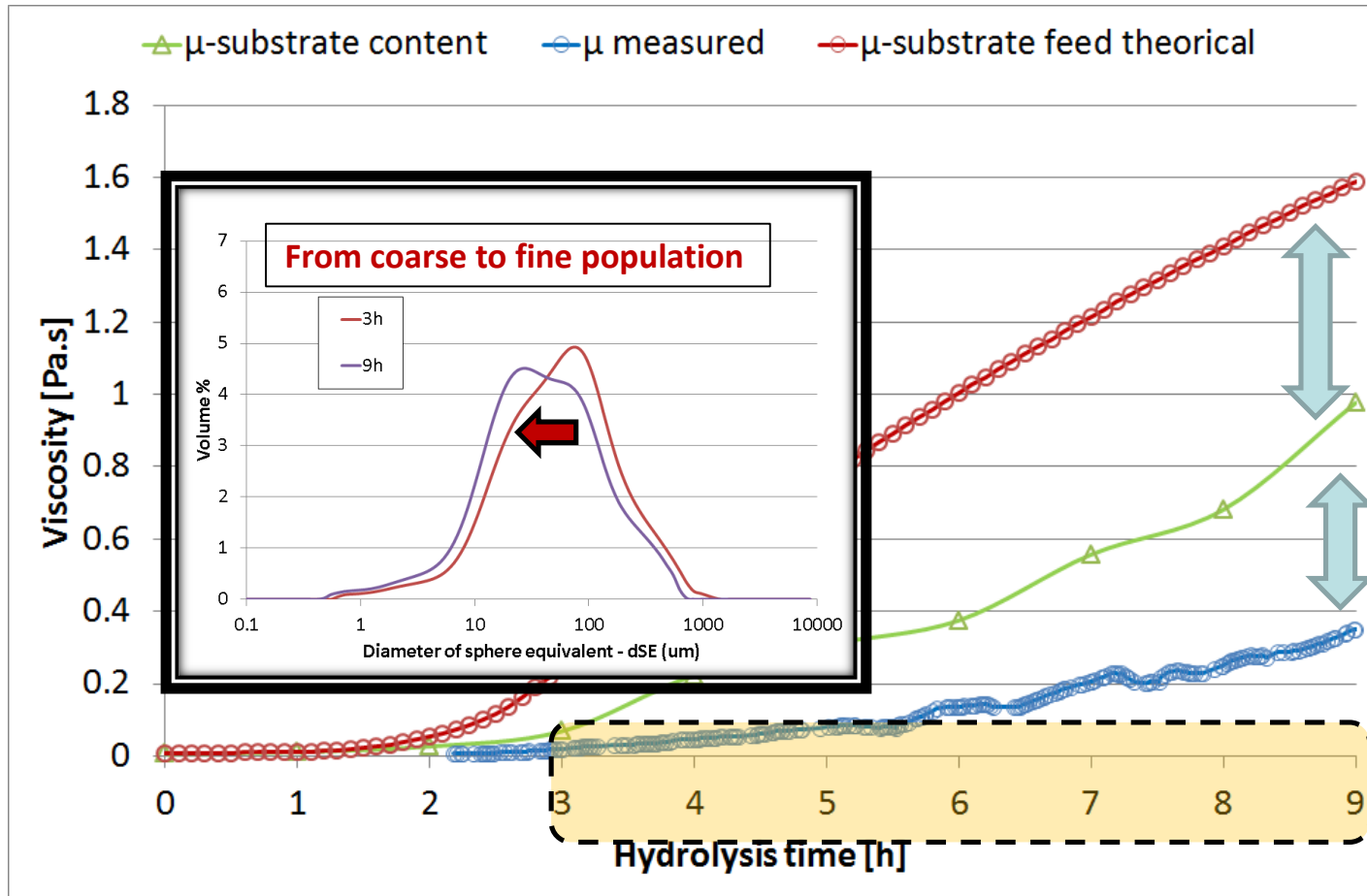
$Q_s = 23.3$ gdm/h

Enzyme: **25**FPU/g cellulose

Phase feeding

Substrate solubilization

Fiber modification

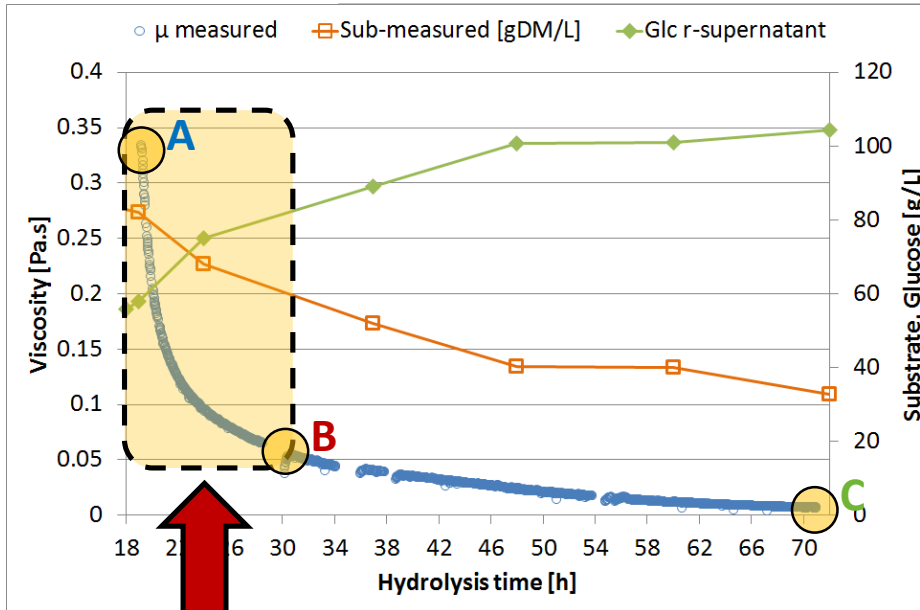


Final [SCB] = **140** gdm/L
 Qs = **23.3** gdm/h
 Enzyme: **25FPU/g** cellulose
Phase feeding

Substrate solubilization

Fiber modification

Mechanism of liquefaction

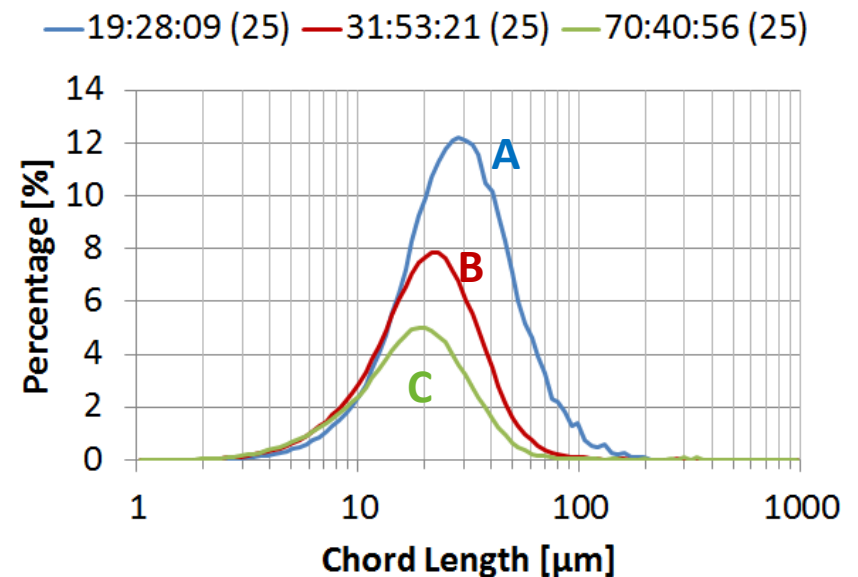


Drastic fall in viscosity

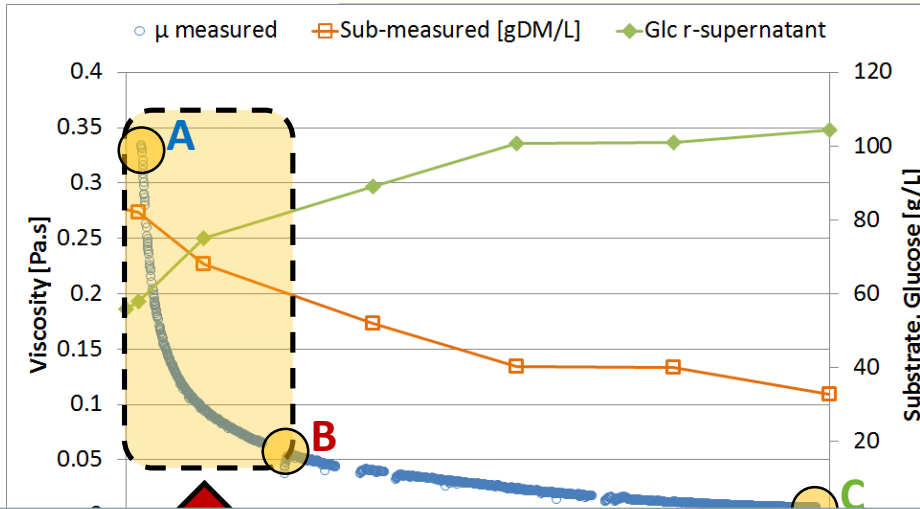
From **A** to **B**: big particles were broken down into smaller particles.

From **B** to **C**: solubilization of small particles

End of feeding
Final [SCB] = **140** gdm/L
Enzyme: **25**FPU/g cellulose



Mechanism of liquefaction



End of feeding
Final [SCB] = **140** gdm/L
Enzyme: **25**FPU/g cellulose

— 19:28:09 (25) — 31:53:21 (25) — 70:40:56 (25)

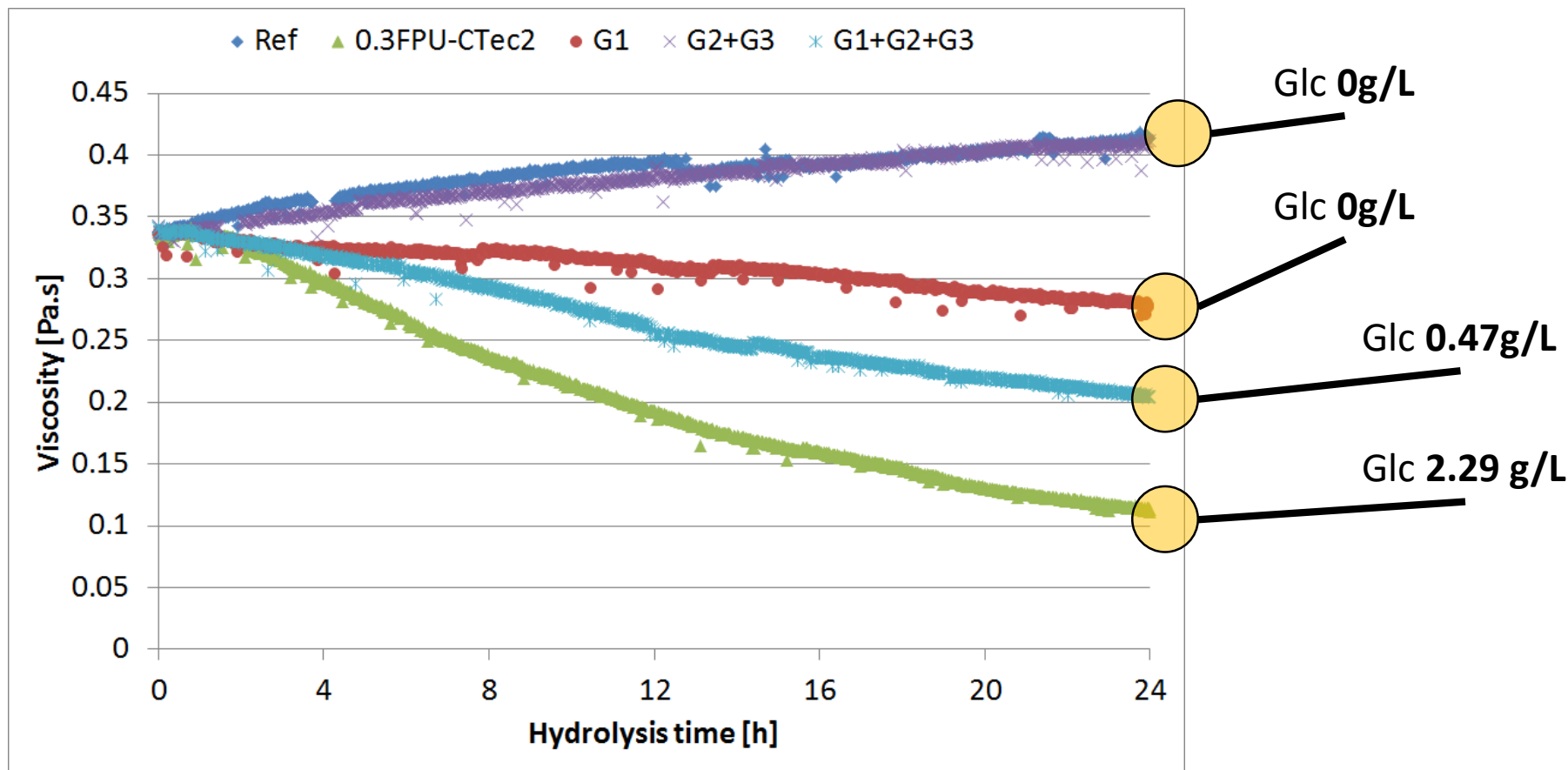
Conclusion 4

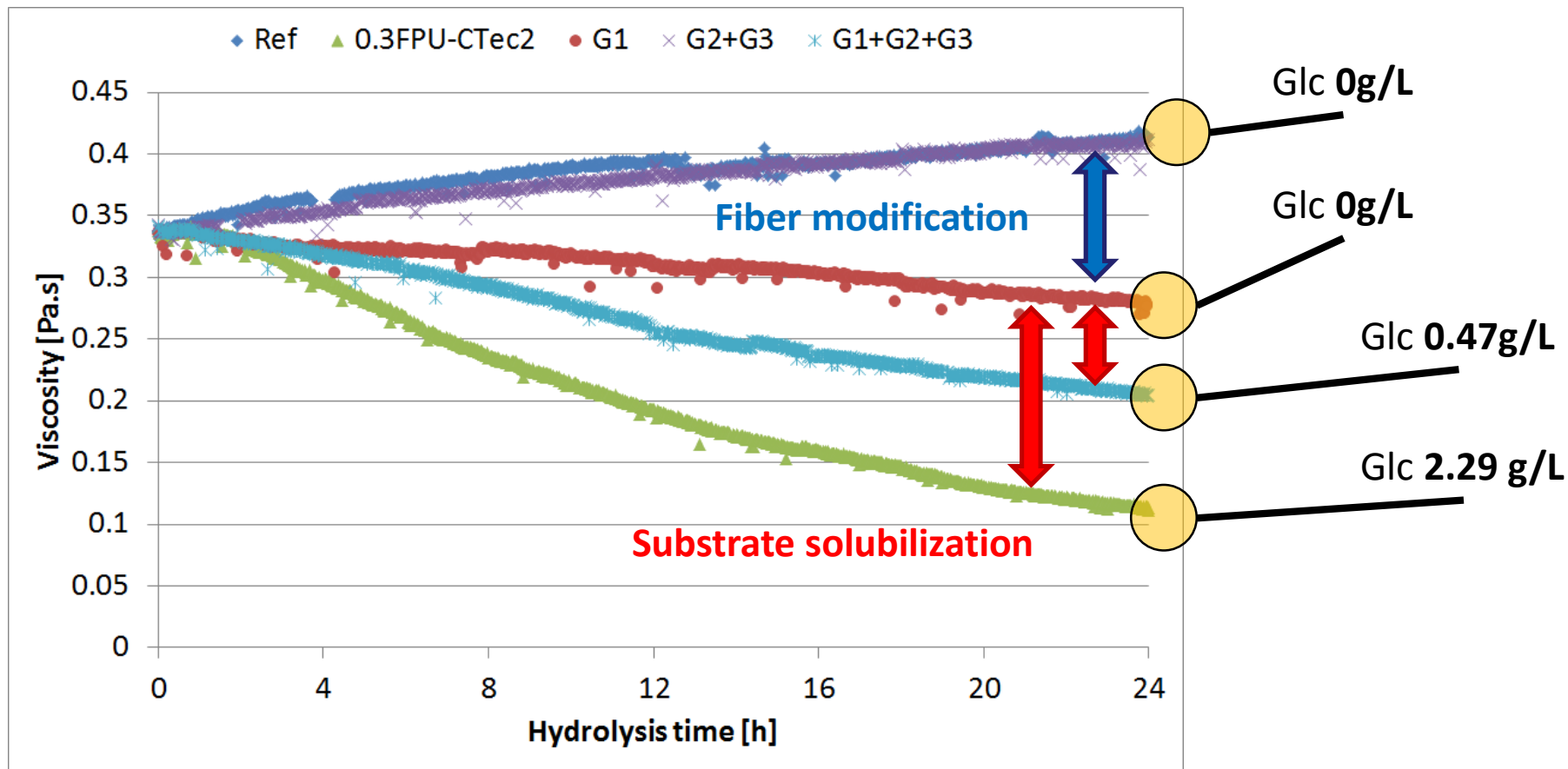
- Enzyme reduces suspension viscosity by two mechanisms: modification of fiber and solubilization of substrate.
- Suspension viscosity was strongly depend on big particles → key to efficient liquefaction.

Chord Length [μ m]

Relationship between physical and biochemical results

Paper pulp 30gdm/L





Conclusions

1. Endo-glucanase = main enzymes in the liquefaction of lignocellulosic suspension.
2. Exo-glucanase and β -glucosidase can improve liquefaction efficiency by synergist with Endo-glucanase.
3. The reduction of suspension viscosity is related to two phenomena:
 - i. Fiber modification,
 - ii. Solubilization of substrate.
4. The viscosity rising step on pretreated sugarcane bagasse is occurred with an evolution of population from fine to coarse. Its mechanism need to be investigated through alternatives experiments and analysis.

Thank for attention!

